

a yellow solid. Recrystallization from THF gave yellow needles: mp 250 °C dec; IR (KBr) 1660, 1615, 1296 cm^{-1} ; ^1H NMR ($\text{DMSO}-d_6$) 3.47 (CH_2 , s, 2 H), 6.70 ($\text{CH}=\text{C}$ —, s, 2 H); ^{13}C NMR ($\text{DMSO}-d_6$) 28.9, 133.5, 146.06, 185.1, 186.8. Anal. Calcd for $\text{C}_{28}\text{H}_{16}\text{O}_8$: C, 70.00; H, 3.36. Found: C, 69.91; H, 3.49.

From 8. The calixarene 8 (1.0 g, 1.36 mmol) was oxidized by a similar method to that described above to yield 0.37 g (57%) of 7.

From 6. The calixarene 6 was oxidized by a similar method to that described above to give 7 in an 84% yield.

Calix[4]hydroquinone (6). 7 (0.6 g, 1.25 mmol) was dispersed in 250 mL of CHCl_3 at 65–70 °C. To the suspension was added a solution of 3.5 g of hydrosulfite in 30 mL of water. The mixture was refluxed vigorously for 2 h at 80–90 °C. After the reaction mixture had been cooled, it was allowed to stand overnight at rt to give a white crude solid. It was recrystallized from $\text{MeOH}-\text{H}_2\text{O}$, which contained a small amount of SnCl_2 and HCl , to yield 0.57 g (93%) of 6 as white plates: mp 400–450 °C dec; IR (KBr) 3200, 1620, 1470 cm^{-1} ; ^1H NMR (acetone- d_6) 3.74 (CH_2 , bs, 2 H), 6.60

(Ar, s, 2 H), 7.82 (OH, s, 1 H), 9.79 (OH, s, 1 H); ^{13}C NMR (acetone- d_6) 32.7, 116.7, 130.6, 143.2, 152.9; MS m/e 489 ($\text{M} + \text{H}$). Anal. Calcd for $\text{C}_{28}\text{H}_{24}\text{O}_8$: C, 68.85; H, 4.95. Found: C, 68.81; H, 4.94.

Acknowledgment. We thank Mr. Y. Akima, Mr. T. Ohyada, and Mr. S. Ohta for carrying out part of the experiments. We are grateful to Dr. Y. Kai of Osaka University for providing the X-ray crystal structure analysis of compound 7 and to Dr. M. Komatsu of Osaka University for providing part of NMR and IR data. We also wish to thank Mr. S. Horiguchi, vice-president of Dainichiseika Color & Chemicals Mfg Co., Ltd., for helpful suggestions on diazo coupling reaction and his support during the course of this work.

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Comparative Studies on the Reactivity of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione, 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione, and Bicyclomycin. Examination of a Key Structural Element Necessary for Bicyclomycin-Mediated Transformations

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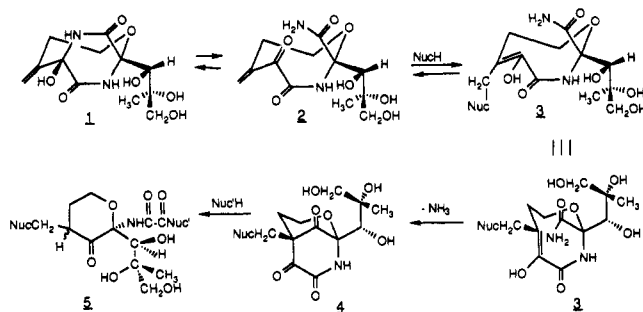
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Two select mimics, 4-methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) and 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) of the structurally novel antibiotic, bicyclomycin (1), have been prepared. Comparison of the chemical reactivity of 7 versus 1 both in the presence and absence of added nucleophiles at various "pH" values has provided important new information concerning the role of key structural elements present in bicyclomycin. The product profiles determined for 7 indicated that modification of the terminal double bond proceeded through an α,β -unsaturated ring imine intermediate (i.e., 43). Correspondingly, activation of the *exo*-methylene group in bicyclomycin is believed to occur through initial hemiaminal bond scission to give a ring-opened α,β -unsaturated carbonyl species (i.e., 2). Functionalization of the terminal double bond in 7 has been shown to proceed under milder conditions than that required for 1. These results demonstrated that incorporation of the *exo*-methylene group within the O(2)–C(3)–C(4)–C(5) bridge in 1 required that the terminal double bond activation pathway proceed by an alternative, energetically more-demanding pathway than that observed for 7. Ramifications of the decreased reactivity noted for 1 are to allow other functional groups (i.e., the C(1)-triol moiety) in the antibiotic to have important catalytic roles in the drug modification processes and to permit thiolate species (the proposed biological targets?) to effectively compete with other nucleophiles for 2.

Bicyclomycin (1) is a structurally unique antibiotic bearing no resemblance to any other known class of antibiotics.^{1–3} The structure and relative configuration of 1 was determined by Tokuma and co-workers by single-crystal X-ray analysis.^{4,5} Subsequently, Maag and his

Scheme I. Proposed Pathway for the Mode of Action of Bicyclomycin (1)



group established the absolute configuration through synthesis and X-ray analysis of a bicyclomycin acid-cata-

(1) For an excellent review, see: Williams, R. M.; Durham, C. A. *Chem. Rev.* 1988, 88, 511.

(2) (a) Miyoshi, T.; Miyairi, N.; Aoki, H.; Kohsaka, M.; Sakai, H.; Imanaka, H. *J. Antibiot.* 1972, 25, 569. (b) Kamiya, T.; Maeno, S.; Hashimoto, M.; Mine, Y. *Ibid.* 1972, 25, 576. (c) Nishida, M.; Mine, Y.; Matsubara, T.; Goto, S.; Kuwahara, S. *Ibid.* 1972, 25, 594.

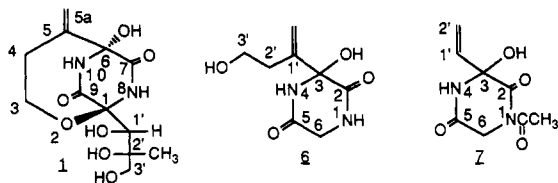
(3) (a) Miyamura, S.; Ogasawara, N.; Otsuka, H.; Niwayama, S.; Tanaka, H.; Take, T.; Uchiyama, T.; Ochiai, H.; Abe, K.; Koizumi, K.; Asao, K.; Matsuki, K.; Hoshino, T. *J. Antibiot.* 1972, 25, 610. (b) Miyamura, S.; Ogasawara, N.; Otsuka, H.; Niwayama, S.; Tanaka, H.; Take, T.; Uchiyama, T.; Ochiai, H. *Ibid.* 1973, 26, 479.

(4) Tokuma, Y.; Koda, S.; Miyoshi, T.; Morimoto, T. *Bull. Chem. Soc. Jpn.* 1974, 47, 18.

lyzed dehydrative rearrangement product.⁶

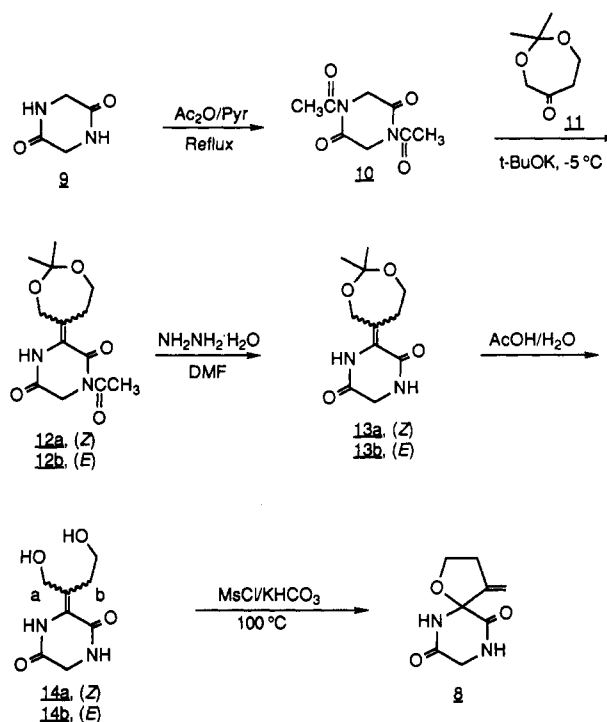
Most proposals have suggested that bicyclomycin reacts with nucleophiles necessary for the remodeling of the peptidoglycan assembly within the bacterial cell wall and that drug binding occurs at the *exo*-methylene group in 1.⁷⁻¹⁴ This hypothesis stems from the pioneering studies of Iseki and co-workers who demonstrated that bicyclomycin reacts with NaSMe at the terminal double bond under basic conditions (pH 12.5).^{7a} Recently, the structural assignment for this methanethiolate adduct has been revised.¹⁵ Additional information concerning the *exo*-methylene modification process was provided by the Williams group through the use of bicyclomycin model compounds.¹⁰ These investigators have concluded that the minimum structural requirements needed in 1 for NaSMe binding under basic conditions included the presence of the following: (1) a free (N-H) amide at N(10); (2) an *exo*-methylene moiety at C(5); (3) a bridgehead hydroxyl at C(6); and (4) a C(1') hydroxyl moiety. In a recent series of papers¹²⁻¹⁵ focusing on the reactivity of bicyclomycin with thiols and amines under near neutral conditions, Kohn and Abuzar have advanced a novel mechanism for the activation of 1 (Scheme I). In this scenario, drug activation occurred by initial ring cleavage of the hemiaminal bond producing 2. Subsequent nucleophilic addition to the α,β -unsaturated carbonyl system generated enol 3, which undergoes an intramolecular mixed-Claisen condensation to yield 4 and NH₃. Incorporated in this hypothesis is the speculation that 4 may serve as an efficient trap for additional nucleophiles present at the drug binding site to give the disubstituted adduct 5.

These combined studies have provided clues on the mode of action of bicyclomycin. An important question that still remains unanswered is what is the biological significance for the placement of the *exo*-methylene moiety within the unique bicyclic ring system present in 1? To provide information concerning this inquiry, we have attempted to compare the reactivity of two bicyclomycin mimics, 3-hydroxy-3-(3-hydroxy-1-methylenepropyl)-2,5-piperazinedione (6) and 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7), with bicyclomycin. In 6 and 7 the

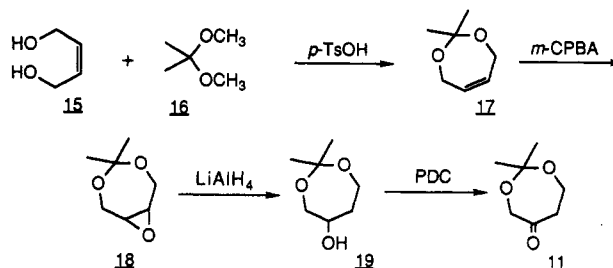


terminal double bond is not constrained within a bicyclic ring system. Our results indicated that alternative pathways for double-bond activation existed in these systems that are not accessible to the *exo*-methylene group in the

Scheme II. Synthetic Sequence for the Preparation of Model Compound 8



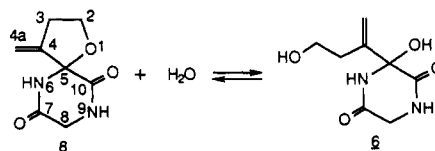
Scheme III. Synthetic Route for the Preparation of Compound 11



antibiotic. The potential significance of this finding in relationship to the mode of action of the drug is discussed.

Results and Discussion

Selection of Compounds. 3-Hydroxy-3-(3-hydroxy-1-methylenepropyl)-2,5-piperazinedione (6) and 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) were chosen to probe the importance of the bridging unit in bicyclomycin-mediated transformations. In both model compounds this structural constraint was deleted. We had anticipated that 6 would be accessible under select hydrolytic conditions through the known 4-methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione⁶ (8). In 7, the N(1)-acetyl group was incorporated within the molecule to aid in the synthesis and isolation of this mimic.



Synthesis of Compounds. The protected bicyclomycin model compound 8 was prepared according to the procedure of Maag and co-workers⁶ with slight modification (Scheme II). Treatment of 2,5-piperazinedione (9) with Ac₂O in the presence of pyridine gave N,N'-diacetyl-2,5-piperazinedione¹⁶ (10). Compound 10 was condensed with

(5) For a related study, see: Kohn, H.; Abuzar, S.; Korp, J. D.; Zektzer, A. S.; Martin, G. E. *J. Heterocycl. Chem.* 1988, 25, 1511.

(6) Maag, H.; Blount, J.; Coffen, D.; Steppe, T.; Wong, F. *J. Am. Chem. Soc.* 1978, 100, 6786.

(7) (a) Someya, A.; Iseki, M.; Tanaka, N. *J. Antibiot.* 1979, 32, 402. (b) Tanaka, N.; Iseki, M.; Miyoshi, T.; Aoki, H.; Imanaka, H. *Ibid.* 1976, 29, 155. (c) Someya, A.; Iseki, M.; Tanaka, N. *Ibid.* 1978, 31, 712.

(8) Williams, R. M.; Armstrong, R. W.; Dung, J.-S. *J. Med. Chem.* 1985, 28, 733.

(9) Pisabarro, A. G.; Canada, F. J.; Vasquez, D.; Arriaga, P.; Rodriguez-Tebar, A. J. *J. Antibiot.* 1986, 34, 914.

(10) (a) Williams, R. M.; Tomizawa, K.; Armstrong, R. W.; Dung, J.-S. *J. Am. Chem. Soc.* 1985, 107, 6419. (b) Williams, R. M.; Tomizawa, K.; Armstrong, R. W.; Dung, J.-S. *Ibid.* 1987, 109, 4028.

(11) Abuzar, S.; Kohn, H. *J. Am. Chem. Soc.* 1988, 110, 4089.

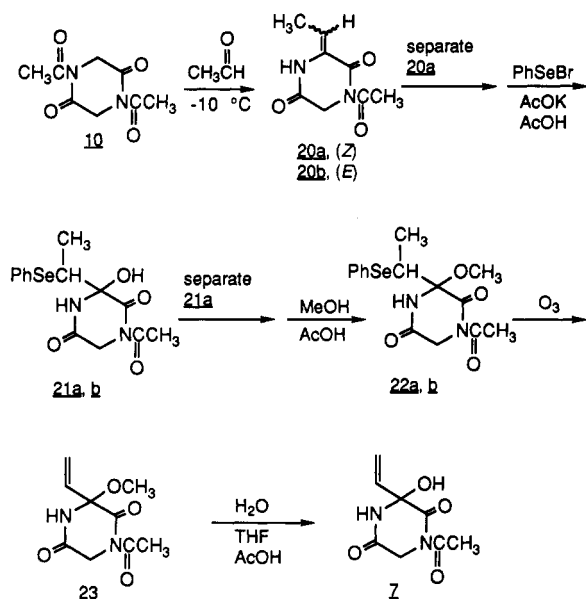
(12) Abuzar, S.; Kohn, H. *J. Am. Chem. Soc.* 1989, 111, 4895.

(13) Abuzar, S.; Kohn, H. *J. Org. Chem.* 1989, 54, 4000.

(14) Abuzar, S.; Kohn, H. *J. Am. Chem. Soc.* 1990, 112, 3114.

(15) Kohn, H.; Abuzar, S. *J. Am. Chem. Soc.* 1988, 110, 3661.

Scheme IV. Synthetic Sequence for the Preparation of Model Compound 7

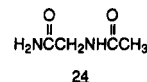


4,4-dimethyl-3,5-dioxacycloheptanone⁶ (11) (tBuOK, DMF, 0 °C) to give 12a and 12b as a 3:2 mixture of the (Z)- and (E)-stereoisomers, respectively. Hydrazinolysis of 12a and b followed by treatment with aqueous acetic acid yielded a 1:1 mixture of diols 14a and 14b. Treatment of the binary mixture of 14a and 14b with mesyl chloride in the presence of KHCO₃ (100 °C) furnished the desired 4-methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) (11% overall yield from 9). Ketone 11 was prepared in four steps from *cis*-1,4-dihydroxy-2-butene (15) (Scheme III, 22% overall yield). Treatment of 15 with 2,2-dimethoxypropane (16) in the presence of a catalytic amount of *p*-toluenesulfonic acid monohydrate gave 17.¹⁷ Epoxidation of 17 with *m*-CPBA followed by reduction with LiAlH₄ produced alcohol 19, which was oxidized with pyridinium dichromate (PDC) in the presence of catalytic amounts of pyridinium trifluoroacetate to yield 11.

The procedure employed for the synthesis of the second bicyclomycin mimic, 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7), is outlined in Scheme IV beginning with 1,4-diacetyl-2,5-piperazinedione (10). Addition of tBuOK in tBuOH to a DMF solution containing 10 and acetaldehyde gave a 7:1 isomeric mixture of (Z)-20a and (E)-20b, 1-acetyl-3-ethylidene-2,5-piperazinedione,¹⁸ respectively (67% yield). Noteworthy, reduction of the temperature for this transformation from the previously suggested value of 0 °C^{19,20} to -10 °C led to an improvement in the yield (37–67%) and the isolation of the minor isomeric product 20b. Treatment of 20a²¹ with PhSeBr and KOAc²² furnished after aqueous workup 21a and 21b

(53% yield) as a 2.8:1 diastereomeric mixture.²³ Dissolution of the major product 21a in methanolic AcOH generated 22a (3%) and 22b (39%) along with recovered starting material (47%). Introduction of the terminal double bond was accomplished by ozonolysis of 22a and 22b to give 23 (88% yield), which was then converted in situ to the target compound 7 by dissolution in an H₂O–AcOH–THF solution (“pH” 2.0–4.0).²⁴

Attempts to isolate purified samples of 7 proved difficult. The C-3 methoxy to C-3 hydroxy interchange process (23 → 7) was readily monitored by ¹³C NMR spectroscopy in a D₂O–DCI–THF-*d*₈ solution. Formation of 7 was accompanied by the disappearance of signals at 51.46 (C(3)–OCH₃) and 87.19 (C(3)) ppm for 23 and the appearance of new peaks at 49.26 and 82.21 ppm for CH₃OD and the C(3) signal in 7, respectively. Correspondingly, use of a D₂O–CD₃CO₂D–THF-*d*₈ (“pD” 2.0) solution containing 23 led to the efficient production (rt, 16 h) of 7 (¹H and ¹³C NMR spectroscopic analyses). High-resolution FAB mass spectral analysis of the product mixture prior to workup confirmed the formation of 7 as the monodeuterio adduct. Neutralization (“pH” 6.9) of half of the reaction mixture followed by removal of the solvent (35 °C) in vacuo led to the production of *N*_α-acetylglycinamide²⁵ (24). Azeotropic



removal of the solvent from the remaining half of the reaction by the successive addition of H₂O gave a crude sample (~70% pure, ¹H and ¹³C NMR analyses) of 7 as an oil which resisted further purification.

Comparative Studies on the Reactivity of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) and Bicyclomycin (1). A tacit assumption in the selection of 4-methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) as a bicyclomycin model compound was that 8 would exist in equilibrium with 3-hydroxy-3-(3-hydroxy-1-methylenepropyl)-2,5-piperazinedione (6) under select conditions. Information concerning this notion and the reactivity of the *exo*-methylene group in 8 was achieved by examining the reactivity of 8 and 1 in H₂O and alcoholic solutions under neutral, acidic, and basic conditions.

Treatment of 8 with EtSH (“pH” 8.0), benzyl mercaptan (“pH” 8.1), and imidazole (“pH” 9.0–11.0) at room temperature in THF–H₂O (3:1) mixtures (48–96 h) led to the recovery of starting material in all cases.²⁶ No evidence for product formation was detected by TLC analyses. This result differed markedly from 1 in which modification of the *exo*-methylene group by the added nucleophile was observed to occur under comparable conditions.^{11–14}

Acid hydrolysis of 8 (0.1 N HCl, 100 °C, 90 min; 1 N H₂SO₄, THF–H₂O (3:1), 80 °C, 4 h) gave after workup 3-hydroxy-4-(2-hydroxyethyl)-2(5*H*)-furanone²⁷ (25) and glycineamide hydrochloride²⁵ (26). Verification of the formation of 25 was accomplished by conversion of this adduct to the diacetate 27²⁷ with Ac₂O and pyridine. Addition of benzyl mercaptan to an acidic (“pH” 1.0)

(16) Abderhalden, E.; Komm, E. Z. *Physiol. Chem.* 1924, 139, 181; *Chem. Abstr.* 1925, 995².

(17) Elliott, W.; Fried, J. J. *Org. Chem.* 1976, 41, 2469.

(18) Distinctive patterns existed in the ¹H NMR spectra for 20a and 20b which allowed their differentiation and identification. Within this pair of compounds a significant deshielding effect (0.24–0.30 ppm) was noted for the methine and methyl protons in closest proximity to the carbonyl group. This pattern has been attributed to the anisotropic effect provided by the C(2)-carbonyl group. Similar trends were noted for comparable pairs of geometric isomers prepared in this study.

(19) Gallina, C.; Liberatori, A. *Tetrahedron Lett.* 1973, 14, 1135.

(20) Shin, C.; Sato, K.; Ohtsuka, A.; Kazutoshi, M.; Yoshimura, J. *Bull. Chem. Soc. Jpn.* 1973, 46, 3876.

(21) Although separation of the isomers is not necessary for the overall preparation of 7 this step was introduced to facilitate the separation of subsequent product mixtures and their spectral identification.

(22) Sharpless, K. B.; Young, M. W. *J. Org. Chem.* 1975, 40, 947.

(23) Treatment of 20a with either PhSeCl or PhSeBr in the presence of silver salts (silver trifluoroacetate or silver acetate) gave (Z)- and (E)-1-acetyl-3-(1-(phenylselenenyl)ethylidene)-2,5-piperazinedione (39–42% yield).

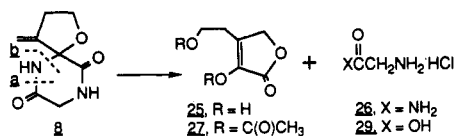
(24) Direct ozonolysis of 21a led to the production of an intimate mixture containing 7 which we were not able to further purify.

(25) The reaction product was compared to an authentic sample purchased from Aldrich Chemical Co.

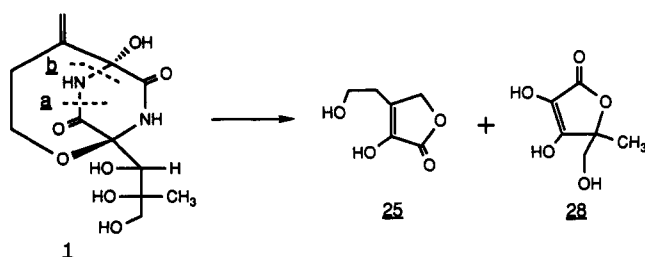
(26) No detectable reactions (NMR analysis) of aqueous solutions of 8 were noted for aqueous solutions maintained at pD 2.3–4.4 (rt, 24 h; 100 °C, 90 min) and at pD 1.0 (rt, 24 h; 50 °C, 12 h).

(27) Kohn, H.; Abuzar, S. *J. Org. Chem.* 1988, 53, 2769.

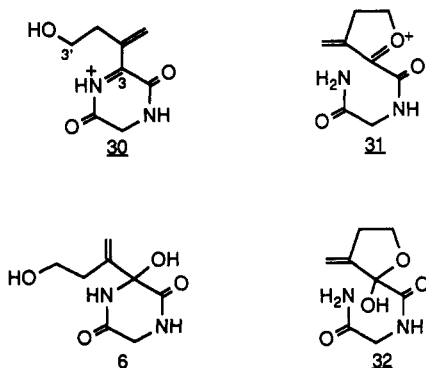
THF-H₂O mixture containing 8 led only to the formation of 25 and 26. No evidence was obtained for the formation of thiolate-substituted products (¹H NMR and TLC analyses).



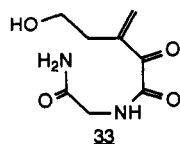
Compound 25 has been isolated along with 28 in the acid hydrolysis (0.1–1.0 N H₂SO₄, 100 °C, 90 min) of bicyclomycin.²⁷ The overall pathway for the conversion of 1 to 25 and 28 was not ascertained in this previous study. Both amide (i.e., route a) and hemiaminal (i.e., route b) bond cleavage pathways were consistent with the reaction products.²⁷ Our observation that 26 is produced together with 25 from 8 supports the notion that in both compounds acid hydrolysis of the piperazinedione ring proceeded by an overall hemiaminal bond scission (route b), since cleavage of 8 by a pathway similar to route a in 1 would have yielded glycine hydrochloride (29) in place of glycine amide hydrochloride (26).



Several pathways are consistent with the hemiaminal bond cleavage process (route b) noted for 8. These differ principally in whether initial C(5)–O(1) or C(5)–N(6) bond scission in 8 occurs to generate 30 and 31, respectively, and whether subsequent reaction with H₂O preferentially proceeds at the terminal end of the conjugated ions 30 and 31 to give the *exo*-methylene-functionalized alcohol or at C(3) in 30 and the corresponding carbon in 31 to give hemiaminal 6 and hemiketal 32, respectively.

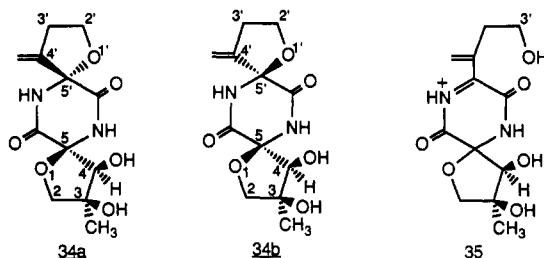


Both 6 and 32 would be expected to furnish the reactive α,β -unsaturated carbonyl intermediate 33 capable of undergoing conjugate addition by H₂O. Suggestive infor-



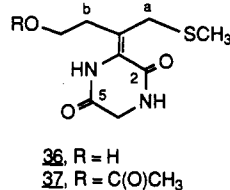
mation that the reaction proceeded by initial C–O bond scission (i.e., 30) was derived from the study of Maag and co-workers.⁶ Treatment of 1 in aqueous H₂SO₄ gave 34a and 34b.⁶ Similarly, dissolution of 1 in 1 N propanolic

H₂SO₄ (98 °C, 90 min) led to the same diastereomeric mixture. Correspondingly, attempted methanolysis (1 N methanolic H₂SO₄, 65 °C, 90 min) or propanolysis (1 N propanolic H₂SO₄, 98 °C, 90 min) of 8 led to the recovery of starting spirofuran in both instances.



These cumulative results provided evidence that C–O bond scission in 1 and 8 is promoted by acid.²⁸ The similar product profiles obtained for 1 and 8 in acidic water and alcoholic solutions suggested that under these conditions both reactions proceeded through a ring-opened intermediate (i.e., 35, 30). The formation of a terminal double bond functionalized adduct (i.e., 25) in water but not in alcoholic solvent systems from 1 and 8 has been attributed in part to the inability of methanol or propanol to effectively compete with the corresponding internal C(3') primary alcohol group in 35 and 30 for the α,β -unsaturated iminium ion system.²⁹ Unfortunately, the absence of methanolic and propanolic products from 8 did not permit us to speculate on whether the *exo*-methylene functionalization process observed for this compound in strong aqueous acid proceeded by the direct addition of H₂O to 30 or through ring-opened intermediate 33.

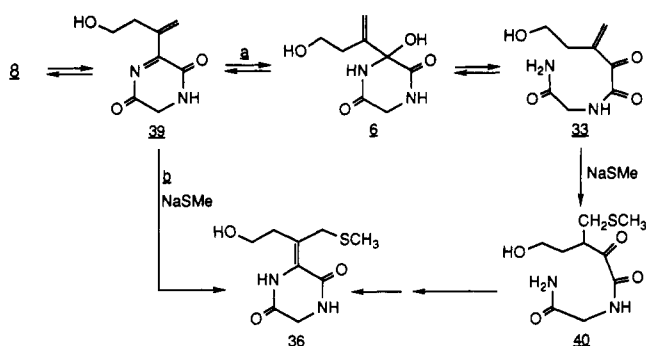
Our study on the chemical reactivity of 4-methylene-1-oxa-6,9-diaspiro[4.5]decane-7,10-dione (8) versus bicyclomycin concluded with the treatment of 8 with NaSMe in base (3:1 THF-H₂O, "pH" 12.5, rt, 2 h). Under these conditions a single product 36 was isolated (69% yield), which has been tentatively assigned as the (*E*)-stereoisomer on the basis of the observed ¹H NMR data. In both 14b (Scheme II) and 36 the C(b) methylene protons appeared upfield (0.18–0.24 ppm) from the corresponding protons in 14a, suggesting that the C(b) methylene protons were trans to the C(2) ring carbonyl group. Acetylation of 36 furnished 3-(3-acetoxy-1-((methylthio)methyl)-propylidene)-2,5-piperazinedione (37).



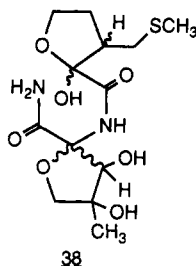
The facility of the NaSMe-mediated reaction with 8 was reminiscent of the reactivity of bicyclomycin with this nucleophile under identical conditions.^{7a,15} In these reactions, functionalization of the *exo*-methylene group proceeded along with cleavage of the piperazinedione ring to generate the diastereomeric C-5a-substituted product 38. Several mechanisms for the conversion of 8 to 36 can be envisioned. Two of these are presented in Scheme V and proceed through 39. In one route, hydration of the imine bond in 39 gives 6 which can undergo ring opening

(28) A similar process is likely to have occurred in the production of 7 from 23.

(29) For the relative nucleophilicities of H₂O versus alcohols, see: (a) Schadt, F. L.; Bentley, T. W.; Schleyer, P. v. R. *J. Am. Chem. Soc.* 1976, 98, 7667. (b) Bentley, T. W.; Schadt, F. L.; Schleyer, P. v. R. *Ibid.* 1972, 94, 993.

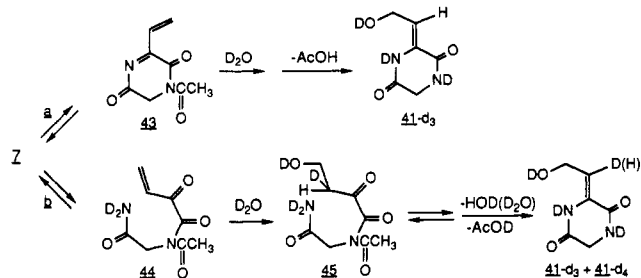
Scheme V. Proposed Pathway for the NaSMe-Mediated Modification of Compound 8 in Base

to give the α,β -unsaturated enone 33. Subsequent thiolate addition followed by ring closure affords 36. In the second pathway depicted, thiolate addition to the α,β -unsaturated system in 39 generates 36 without the intervention of 6 and 33. Information related to this transformation was provided by the treatment of both 8 and 1 with NaSMe in methanolic 0.03 N NaOMe ("pH" 11.9). With 8 no reaction products were detected (rt, 24 h), while 1 furnished the diastereomeric C-5a-substituted products¹⁵ 38a and 38b in 66% yield (rt, 2 h). These results suggested that under aqueous basic conditions conversion of 8 to 33 via 6 may have occurred prior to the addition of NaSMe to the *exo*-methylene group. Alternatively, decreased amounts of 39 may have been generated in the basic methanol solutions versus the corresponding aqueous solutions, thereby diminishing the likelihood of the conjugate addition of NaSMe to the α,β -imine system.³⁰ Attempts to gain additional information concerning these mechanistic notions by examining the reactivity of the bis-spiro bicyclic adducts 34a and 34b⁶ under comparable conditions were unsuccessful. Treatment of either a 1:1 THF-H₂O mixture ("pH" 12.4, rt, 24 h) or a methanolic 0.03 N NaOMe ("pH" 11.8, rt, 24 h) solution containing 34a,b with NaSMe (2 equiv) led to the generation of a complex mixture in both cases. Careful preparative TLC of the aqueous THF reaction permitted the isolation of the major component in the product mixture. ¹H and ¹³C NMR analysis indicated that this compound corresponded to one of the two starting diastereomers 34a or 34b, suggesting that these isomers displayed differential reactivities under basic conditions.



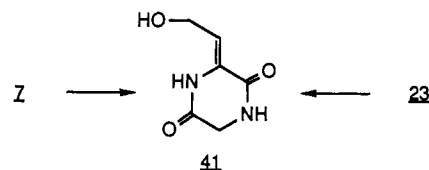
The lack of reactivity of 8 under moderate reaction conditions prevented us from addressing the chemical ramifications of incorporating the *exo*-methylene group in the constrained O(2)-C(5) bridge in bicyclomycin. Pertinent information concerning this issue was provided by the second model compound 7.

Comparative Studies on the Reactivity of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) and Bicyclomycin (1). The inability to modify the *exo*-

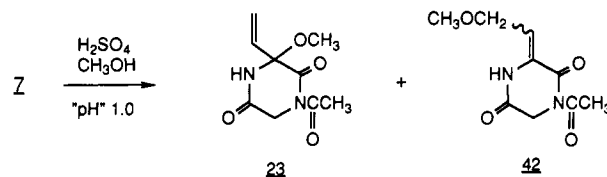
Scheme VI. Potential Mechanisms for the Conversion of Compound 7 to Compound 41 in D₂O Solutions

methylene group in 8 under moderate conditions has been attributed in part to the ease of spiro-ring formation from the corresponding α,β -unsaturated iminium ion (i.e., 30) or imine (i.e., 39) system. In the simplified model compound 7, spiro-ring formation is not possible prior to conjugate addition to the terminal double bond. Furthermore, incorporation of the C(3)-hydroxy group in 7 ensured that the reaction proceeded through a hemiaminal species. These modifications permitted us to compare the reactivity of this compound with bicyclomycin at intermediate "pH" values as well as in acid and base.

Compound 7 was generated from 23 in aqueous acid ("pH" 2-4). At these "pH" values 7 was stable for extended periods of time (>2 d) at room temperature. Reduction of the "pD" of the solution to 1.0 led to the formation of 3-(2-hydroxyethylidene)-2,5-piperazinedione (41). A similar result was observed for 23 at this "pD" value. Both transformations took approximately 16 h to complete at room temperature and proceeded with the loss of the N(1)-acetyl group.



A parallel reaction for 7 was performed in MeOH. Treatment of 7 with methanolic H₂SO₄ ("pH" 1.0, rt, 10 h) gave 23 as the major product (53% yield). A minor adduct (2% yield) was also isolated which has been tentatively assigned as the (*E*)- and (*Z*)-isomers of 42 (¹H NMR analysis).³¹



Two likely pathways for the conversion of 7 to 41 in acid are presented in Scheme VI. These routes differ in whether a ring-cleavage process occurred. Information in support of pathway a was derived from the ¹H NMR spectrum of 41 obtained from the acid hydrolysis experiment conducted in D₂O (pD 1.0), which demonstrated that no deuterium incorporation had occurred at the olefinic position (δ 5.71). This observation was reinforced by the high-resolution mass spectrum of this adduct, which displayed a prominent peak for the nondeuterated molecular ion after workup. The absence of deuterium in 41 eliminated route b in Scheme VI. This pathway required that

(30) For an example of the addition of thiols to α,β -unsaturated imines, see: Buckwell, S.; Page, M. I.; Longridge, J. L. *J. Chem. Soc., Chem. Commun.* 1986, 1039.

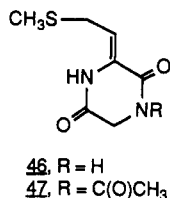
(31) Attempts to confirm the identity of 42 by preparing an authentic sample from 3-methoxypropionaldehyde and 10 with tBuOK/tBuOH were unsuccessful.

a substantial amount of deuterium be incorporated at the vinylic position.³²

Elevation of the "pD" from the conditions employed for the preparation of 7 led to the gradual activation of the terminal double bond in the model compound. Addition of 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) to an aqueous "pD" 6.0 solution (3 h) furnished 41. Correspondingly, dissolution of 7 in a "pD" 8.8 solution (30 min) gave a mixture of 24 (45%) and 41 (13%), respectively. ¹H NMR analyses of 41 isolated from both of these reactions indicated that no deuterium incorporation had occurred at the olefinic site suggesting that a mechanism comparable to Scheme VI route a was operative in these transformations as well. The formation of significant amounts of *N*_α-acetylglycinamide (24) at "pD" 8.8 may signal that competing reaction processes are occurring at higher "pD" values.

The lability of 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) under moderate conditions contrasted to that observed for bicyclomycin (1). Dissolution of 1 at "pD" 5.7–10.2 in the absence of external nucleophiles led to little drug modification (24 h) (NMR analysis). At "pD" 5.7 in a buffered D₂O–THF-*d*₈ (2:1) solution no change in 1 was noted, while at "pD" 10.2 approximately 10% of bicyclomycin had been converted to an unidentified adduct(s).

Efforts to add a sulfur nucleophile to 7 under near-neutral pH conditions proved unsuccessful. Addition of NaSMe to a 3:1 THF–D₂O solution ("pD" 7.0) containing 7 gave only 41 after workup. No evidence was obtained for 3-(2-(methylthio)ethylidene)-2,5-piperazinedione (46) or the corresponding *N*-acetyl derivative 47. We suspect under these conditions the methanethiol was insufficiently ionized³³ to effectively compete with the bulk solvent for the α,β-unsaturated imine system in 43.³⁴



Conclusions

The enhanced reactivity of the terminal double bond in 1-acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) versus bicyclomycin (1) toward nucleophilic attack at moderate "pH" values has been attributed to the differential pathways for activation that exist for these compounds. In 7 nucleophilic addition is promoted by ring imine formation, which permits the terminal double bond to be in conjugation with the endocyclic imine bond (Scheme VI, route a). A similar process for 1, however, is not likely. In bicyclomycin generation of a bridgehead imine bond would be accompanied by a significant amount of ring strain.³⁵ Moreover, this process would not permit the imine bond to be in conjugation with the *exo*-methylene group due to

the structural constraints imposed by the [4.2.2] bicyclic ring system. Accordingly, activation of the terminal double bond in bicyclomycin must proceed by an alternative, energetically more-demanding pathway than that accessible to 7. Previous studies have demonstrated that this process occurs by initial hemiaminal bond cleavage to generate the reactive ring-opened α,β-unsaturated carbonyl species 2.^{11–15} Ramifications of this decreased reactivity for 1 are as follows: (1) to permit other functional groups (i.e., C(1)-triol moiety) present in the antibiotic to play important catalytic roles in the drug activation and binding processes^{10–14} and (2) to necessitate that the basicity of the medium be sufficiently high to activate the *exo*-methylene group in 1. Elevation of the "pH" in this manner should permit key protein thiolate species (i.e., the biological target?) to effectively compete with nucleophiles present in the biological milieu for the presumed reactive α,β-unsaturated carbonyl intermediate 2.

Experimental Section

General Methods. The experimental procedures used in this study were identical to those employed in previous investigations.^{11–15,27} Generous supplies of bicyclomycin were obtained from Fujisawa Pharmaceutical Co., Ltd., Japan. *cis*-1,4-Dihydroxy-2-butene (98%) was purchased from American Tokyo Kasei, Portland, OR. THF and Et₂O were distilled from Na⁰ and benzophenone, and CH₂Cl₂ was distilled from P₂O₅. DMF was fractionally distilled from CaSO₄ under reduced pressure. Pyridine was dried by refluxing with KOH pellets. The pD of the solution was obtained by using the following relationship: pD = pH meter reading + 0.4.³⁶

Preparation of 4,4-Dimethyl-3,5-dioxacycloheptanol (19). To a suspension of LiAlH₄ (10.00 g, 238 mmol) in anhydrous Et₂O (500 mL) was added epoxide 18¹⁷ (24.00 g, 164 mmol) in anhydrous Et₂O (200 mL) dropwise (3 h) at 0 °C. The mixture was allowed to warm to rt and stirred (24 h). The excess LiAlH₄ was destroyed by the successive addition of H₂O (10 mL), an aqueous 15% NaOH solution (10 mL), and H₂O (30 mL).³⁷ The resulting mixture was filtered through a Celite bed, and the filtrate was washed with an aqueous saturated NaCl solution (200 mL), dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (50% ethyl acetate–hexane) to give 19 as a liquid: yield 16.70 g (68%); *R*_f 0.35 (50% ethyl acetate–hexane); IR (neat) 3380, 1385, 1375, 1215, 1155, 1100, 1080, 1045 cm^{−1}; ¹H NMR (CDCl₃) δ 1.23, 1.25 (2 s, 6 H, C(CH₃)₂), 1.62–1.73 (m, 1 H, OCH₂CHH'), 1.73–1.86 (m, 1 H, OCH₂CHH'), 3.42 (br s, 1 H, OH), 3.54–3.84 (m, 5 H, CH, OCH₂CH, OCH₂CH₂); ¹³C NMR (CDCl₃) 24.40 (C(CH₃)₂), 37.50 (OCH₂CH₂), 56.84 (OC–H₂CH₂), 64.98 (OCH₂CH), 67.77 (CH), 100.95 (C(CH₃)₂) ppm; the ¹³C NMR assignments were confirmed by the corresponding APT experiment; MS *m/e* (rel intensity) 146 (3), 131 (52), 115 (11), 101 (21), 75 (100).

Anal. Calcd for C₇H₁₄O₃: C, 57.51; H, 9.65. Found: C, 57.39; H, 9.45.

Preparation of 4,4-Dimethyl-3,5-dioxacycloheptanone (11).⁶ To a CH₂Cl₂ solution (200 mL) of 19 (16.00 g, 110 mmol) were added pyridinium dichromate (84.00 g, 223 mmol) and pyridinium trifluoroacetate (8.47 g, 44 mmol), and then the suspension was mechanically stirred at rt (24 h). The solid was filtered through a silica gel bed, and the solvent was evaporated in vacuo. The residue was purified by flash column chromatography (35% ethyl acetate–hexane) to give 11 as a low-boiling liquid: yield³⁸ 7.38 g (47%); *R*_f 0.78 (50% ethyl acetate–hexane); ¹H NMR (CDCl₃) δ 1.45 (s, 6 H, C(CH₃)₂), 2.81 (t, *J* = 7.0 Hz, 2 H, OCH₂CH₂), 3.94 (t, *J* = 7.0 Hz, 2 H, OCH₂CH₂), 4.00 (s, 2 H, OCH₂C=O); ¹³C NMR (CDCl₃) 24.84 (C(CH₃)₂), 41.63 (OC–H₂CH₂), 56.54 (OCH₂CH₂), 68.49 (OCH₂C=O), 102.11 (C(CH₃)₂), 210.99 (C=O) ppm.

(36) Bates, R. G. *Determination of pH: Theory and Practice*, 2nd ed.; Wiley: New York, 1973; pp 375–376.

(37) Micovic, V. M.; Mihailovic, M. L. *J. Org. Chem.* 1953, 18, 1190.

(38) The volatility of the product did not permit the complete removal of EtOAc. The yield was estimated by ¹H NMR spectroscopy.

(32) The possibility that the deuterium incorporated at the vinylic site in 41 was lost by an enamine H/D exchange process during the chromatographic workup with 20% MeOH–CHCl₃ has been ruled out. Repeated PTLC separation of 20a with 20% MeOD–CHCl₃ led to no detectable deuterium incorporation at the olefinic site.

(33) Crampton, M. R. In *The Chemistry of the Thiol Group*; Patai, S., Ed.; Wiley: London, 1974; pp 379–415.

(34) In the thiol addition studies reported by Page and co-workers,³⁰ higher thiol to imine ratios were utilized. Moreover, this study was conducted in water.

(35) Maier, W. F.; Schleyer, P. v. R. *J. Am. Chem. Soc.* 1981, 103, 1891 and references cited therein.

Preparation of 1-Acetyl-3-(4,4-dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (12a and 12b).⁶ To a solution of 10¹⁸ (9.63 g, 48.6 mmol) and 11 (7.00 g, 48.6 mmol) in dry DMF (100 mL) was added a solution of tBuOK (5.46 g, 48.6 mmol) in tBuOH (100 mL) dropwise (1 h) at -10 °C. The temperature was kept at -10 °C (2 h), and then the solution was warmed to rt and stirred (18 h). The solution was poured into H₂O, neutralized with AcOH, and extracted with CHCl₃ (3 × 200 mL). The organic layers were combined, washed with an aqueous saturated NaCl solution (3 × 200 mL), dried, and concentrated in vacuo. The products were purified by flash chromatography (50% ethyl acetate-hexane) to give a mixture of 12a and 12b: yield 7.54 g (55%). The binary mixture (300 mg) was separated by PTLC (50% ethyl acetate-hexane) to give pure 12a and 12b.

(Z)-1-Acetyl-3-(4,4-dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (12a): yield 152 mg (51%); *R*_f 0.47 (50% ethyl acetate-hexane); IR (KBr) 1715, 1694, 1633, 1360, 1224, 1197, 1076 cm⁻¹; ¹H NMR (CDCl₃) δ 1.42 (s, 6 H, C(CH₃)₂), 2.58 (s, 3 H, CH₃C=O), 3.24 (t, *J* = 6.3 Hz, 2 H, OCH₂CH₂), 3.86 (t, *J* = 6.3 Hz, 2 H, OCH₂CH₂), 4.40 (s, 2 H, CH₂N), 4.41 (s, 2 H, OCH₂C=C), 9.19 (br s, 1 H, NH); ¹³C NMR (CDCl₃) 24.31 (C-(CH₃)₂), 26.98 (CH₃C=O), 29.44 (OCH₂CH₂), 45.13 (CH₂N), 61.18 (OCH₂CH₂ and OCH₂C=C), 102.31 (C(CH₃)₂), 120.03 (CH₂C=C), 141.40 (C=CN), 161.60 (C=CC=O), 165.87 (CH₂C=O), 172.08 (CH₂C=O) ppm; MS (EI) *m/e* (rel intensity) 282 (1), 264 (6), 251 (4), 240 (1), 224 (62), 182 (100); MS (+CI) *m/e* (rel intensity) 283 (1), 265 (4), 225 (100), 183 (8); *M*_r (EI) 282.12100 (calcd for C₁₃H₁₈N₂O₅, 282.12157).

Anal. Calcd for C₁₃H₁₈N₂O₅·H₂O: C, 52.02; H, 6.72; N, 9.33. Found: C, 52.39; H, 6.71; N, 8.98.

(E)-1-Acetyl-3-(4,4-dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (12b): yield 113 mg (38%); *R*_f 0.42 (50% ethyl acetate-hexane); ¹H NMR (CDCl₃) δ 1.41 (s, 6 H, C(CH₃)₂), 2.57 (s, 3 H, CH₃C=O), 2.83 (t, *J* = 6.3 Hz, 2 H, OCH₂CH₂), 3.85 (t, *J* = 6.3 Hz, 2 H, OCH₂CH₂), 4.38 (s, 2 H, CH₂N), 4.74 (s, 2 H, OCH₂C=C), 9.44 (br s, 1 H, NH); ¹³C NMR 24.29 (C(CH₃)₂), 26.83 (CH₃C=O), 29.91 (OCH₂CH₂), 45.16 (CH₂N), 60.28 (OCH₂CH₂), 62.97 (OCH₂C=C), 102.18 (C(CH₃)₂), 119.55 (OCH₂C=C), 143.04 (C=CN), 164.86 (C=CC=O), 166.00 (CH₂C=O), 172.11 (CH₂C=O) ppm; MS (EI) *m/e* (rel intensity) 282 (1), 264 (2), 251 (1), 240 (1), 224 (57), 182 (100); MS (+CI) *m/e* (rel intensity) 283 (9), 265 (5), 225 (100), 183 (19); *M*_r (EI) 282.12100 (calcd for C₁₃H₁₈N₂O₅, 282.12157).

Anal. Calcd for C₁₃H₁₈N₂O₅: C, 55.32; H, 6.38; N, 9.93. Found: C, 54.97; H, 6.49; N, 9.75.

Preparation of 3-(4,4-Dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (13a and 13b).⁶ To a solution of 12a and 12b (6.70 g, 23.8 mmol) in dry DMF (50 mL) was added N₂H₄·H₂O (2.38 g, 47.5 mmol). The solution was stirred at rt (2 h) and then poured into ice-H₂O. The solid was collected and purified by flash column chromatography (10% MeOH-CHCl₃) to give a 3:2 mixture of 13a and 13b: yield 5.22 g (92%); mp 207–209 °C. The following spectral properties have been assigned to 13a and 13b.

(Z)-3-(4,4-Dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (13a): ¹H NMR (DMSO-*d*₆) δ 1.29 (s, 6 H, C(CH₃)₂), 3.14 (t, *J* = 6.1 Hz, 2 H, OCH₂CH₂), 3.65 (t, *J* = 6.1 Hz, 2 H, OCH₂CH₂), 3.79 (s, 2 H, CH₂N), 4.28 (s, 2 H, OCH₂C=C), 8.11, 9.58 (s, 2 H, 2 × NH); ¹³C NMR (DMSO-*d*₆) 24.86 (C(CH₃)₂), 28.44 (OCH₂CH₂), 44.44 (CH₂N), 60.58, 60.85 (OCH₂CH₂ and OCH₂C=C), 104.89 (C(CH₃)₂), 121.16 (CH₂C=C), 132.94 (C=CN), 162.11 (C=CC=O), 165.66 (CH₂C=O) ppm.

(E)-3-(4,4-Dimethyl-3,5-dioxacycloheptylidene)-2,5-piperazinedione (13b): ¹H NMR (DMSO-*d*₆) δ 1.28 (s, 6 H, C(CH₃)₂), 2.62 (t, *J* = 6.1 Hz, 2 H, OCH₂CH₂), 3.68 (t, *J* = 6.1 Hz, 2 H, OCH₂CH₂), 3.81 (s, 2 H, CH₂N), 4.61 (s, 2 H, OCH₂C=C), 8.06, 9.58 (s, 2 H, 2 × NH); ¹³C NMR (DMSO-*d*₆) 24.86 (C(CH₃)₂), 29.10 (OCH₂CH₂), 44.44 (CH₂N), 60.32, 61.51 (OCH₂CH₂ and OCH₂C=C), 104.89 (C(CH₃)₂), 120.90 (CH₂C=C), 133.07 (C=CN), 161.84 (C=CC=O), 165.92 (CH₂C=O) ppm.

Preparation of 3-(3-Hydroxy-1-(hydroxymethyl)propylidene)-2,5-piperazinedione (14a and 14b).⁶ A solution of 13a and 13b (4.80 g, 20 mmol) in 50% aqueous AcOH (100 mL) was stirred at rt (2 h). The solvent was removed in vacuo, and the residue was purified by flash chromatography (methanol-acetone-chloroform (1:7:8)) to give a 1:1 mixture of 14a and 14b:

yield 3.36 g (84%); mp 166–169 °C; *R*_f 0.22 (methanol-acetone-chloroform (1:7:8)); MS (EI) *m/e* (rel intensity) 200 (3), 154 (16), 139 (100), 124 (39), 111 (34). The following spectral properties have been assigned to 14a and 14b.

(Z)-3-(3-Hydroxy-1-(hydroxymethyl)propylidene)-2,5-piperazinedione (14a): ¹H NMR (DMSO-*d*₆) δ 2.73 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂), 3.61 (t, *J* = 6.0 Hz, 2 H, HOCH₂CH₂), 3.82 (s, 2 H, CH₂N), 4.18 (s, 2 H, HOCH₂C=C), 4.65 (br s, 1 H, HOCH₂CH₂), 5.51 (br s, 1 H, HOCH₂C=C), 8.15 (s, 1 H, CH₂NH), 9.54 (br s, 1 H, NHC=C) [The ¹H NMR assignments were confirmed by the corresponding COSY experiment. The assignments for the amide NH and CH₂N proton signals are uncertain and may correspond to the resonances for the corresponding protons in 14b]; ¹³C NMR (DMSO-*d*₆) 32.69 (OCH₂CH₂), 44.30 (CH₂N), 60.51 (OCH₂CH₂), 61.82 (OCH₂C=C), 123.98 (CH₂C=C), 130.39 (C=CN), 161.31 (C=CC=O), 164.69 (CH₂C=O) ppm. The ¹³C NMR assignments were confirmed by the ¹H-¹³C heteronuclear chemical shift connectivity experiment. The assignments for the CH₂N, C=C, and C=O carbon signals are uncertain and may correspond to the resonances for the corresponding carbons in 14b.

(E)-3-(3-Hydroxy-1-(hydroxymethyl)propylidene)-2,5-piperazinedione (14b): ¹H NMR (DMSO-*d*₆) δ 2.49 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂), 3.52 (m, 2 H, HOCH₂CH₂), 3.84 (s, 2 H, CH₂N), 4.50 (d, 1 H, *J* = 4.5 Hz, HOCH₂C=C), 4.80 (t, 1 H, *J* = 4.5 Hz, HOCH₂C=C), 5.12 (br s, 1 H, HOCH₂CH₂), 8.17 (s, 1 H, CH₂NH), 9.75 (br s, 1 H, NHC=C) [The ¹H NMR assignments were confirmed by the corresponding COSY experiment. The assignments for the amide NH and CH₂N proton signals are uncertain and may correspond to the resonances for the corresponding protons in 14a]; ¹³C NMR (DMSO-*d*₆) 31.67 (OCH₂CH₂), 44.42 (CH₂N), 59.18 (OCH₂C=C), 60.00 (OCH₂CH₂), 124.07 (CH₂C=C), 133.38 (C=CN), 161.94 (C=CC=O), 165.64 (CH₂C=O) ppm. The ¹³C NMR assignments were confirmed by the ¹H-¹³C heteronuclear connectivity experiment. The assignments for the CH₂N, C=C, and C=O carbon signals are uncertain and may correspond to the resonances for the corresponding carbons in 14a.

Preparation of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8).⁶ To a 1,4-dioxane solution (150 mL) containing 14a and 14b (2.50 g, 12.5 mmol) were added methanesulfonyl chloride (2.18 g, 18.9 mmol) and KHCO₃ (1.50 g, 15 mmol). The suspension was heated to reflux (15 h), and then the solvent was removed in vacuo, and the residue was purified by flash column chromatography (10% MeOH-CHCl₃) to give 8: yield 591 mg (26%); *R*_f 0.53 (10% MeOH-CHCl₃); mp 211–213 °C; IR (KBr) 3330, 3180, 3070, 2940, 1660, 1650, 1440, 1095, 918 cm⁻¹; ¹H NMR (DMSO-*d*₆) δ 2.60–2.73 (m, 2 H, OCH₂CH₂), 3.75–3.94 (m, 4 H, OCH₂CH₂, CH₂N), 5.05 (s, 1 H, C=CHH), 5.24 (s, 1 H, C=CHH), 8.15, 8.88 (2 s, 2 H, 2 × NH); ¹³C NMR (DMSO-*d*₆) 32.09 (OCH₂CH₂), 44.49 (CH₂N), 65.96 (OCH₂CH₂), 88.78 (OCN), 109.78 (C=CH₂), 148.53 (C=CH₂), 165.51 (2 × C=O) ppm; MS (EI) *m/e* (rel intensity) 182 (2), 154 (15), 139 (100), 125 (4), 111 (67), 98 (55), 81 (39), 67 (80), 53 (73); *M*_r (EI) 182.06945 (calcd for C₈H₁₀N₂O₃, 182.06914).

Preparation of 1-Acetyl-3-ethylidene-2,5-piperazinedione (20a and 20b).¹⁸ To a solution of 10 (10.00 g, 51 mmol) and acetaldehyde (22.25 g, 510 mmol) in dry DMF (100 mL) was added a solution of tBuOK (5.67 g, 51 mmol) in tBuOH (100 mL) dropwise (1 h) at -10 °C. The temperature was kept at -10 °C (1 h), and then the solution was warmed to rt. The solution was poured into H₂O, neutralized with AcOH, and extracted with CHCl₃ (3 × 300 mL). The organic layers were combined, washed with an aqueous saturated NaCl solution (300 mL), dried, and concentrated in vacuo. The residue was purified by flash column chromatography (50% ethyl acetate-hexane) to give 20a and 20b.

(Z)-1-Acetyl-3-ethylidene-2,5-piperazinedione (20a): yield 5.42 g (59%); mp 184 °C (lit.²⁰ mp 182–184 °C); *R*_f 0.69 (50% ethyl acetate-hexane); ¹H NMR (DMSO-*d*₆) δ 1.82 (d, *J* = 7.5 Hz, 3 H, CHCH₃), 2.48 (s, 3 H, CH₃C=O), 4.27 (s, 2 H, CH₂N), 6.15 (q, *J* = 7.5 Hz, 1 H, CHCH₃), 10.25 (s, 1 H, NH); ¹³C NMR (DMSO-*d*₆) 11.81 (CHCH₃), 26.82 (CH₃C=O), 45.90 (CH₂N), 117.49 (CH₃CH=C), 128.51 (CH₃CH=C), 160.42 (C=CC=O), 163.64 (CH₂C=O), 172.13 (CH₃C=O) ppm. The ¹³C NMR assignments were confirmed by the corresponding APT experiment.

(*E*)-1-Acetyl-3-ethylidene-2,5-piperazinedione (**20b**): yield 0.74 g (8%); mp 183–184 °C; R_f 0.73 (50% ethyl acetate–hexane); ^1H NMR (DMSO- d_6) δ 2.06 (d, J = 7.5 Hz, 3 H, CHCH₃), 2.51 (s, 3 H, CH₃C=O), 4.21 (s, 2 H, CH₂N), 5.85 (q, J = 7.5 Hz, 1 H, CHCH₃), 10.36 (s, 1 H, NH); ^{13}C NMR (DMSO- d_6) 13.72 (CHCH₃), 26.98 (CH₃C=O), 45.71 (CH₂N), 122.32 (CH₃CH=C), 127.60 (CH₃CH=C), 162.40 (C=CC=O), 165.50 (CH₂C=O), 171.68 (CH₃C=O) ppm. The ^{13}C NMR assignments were confirmed by the corresponding APT experiment.

Preparation of 1-Acetyl-3-hydroxy-3-(1-(phenylselenenyl)ethyl)-2,5-piperazinedione (21a and 21b). To a suspension of PhSeBr (1.79 g, 7.58 mmol) and KOAc (1.00 g, 10.19 mmol) in AcOH (40 mL) was added **20a** (1.00 g, 5.49 mmol) at rt under N₂. The reaction mixture was stirred at rt (12 h) and then poured into H₂O (100 mL) and extracted with CHCl₃ (3 \times 100 mL). The organic layers were combined and washed with an aqueous saturated NaHCO₃ solution (3 \times 100 mL) and an aqueous saturated NaCl solution (150 mL). The solvent was removed in vacuo, and the residue was purified by flash column chromatography (60% ethyl acetate–hexane) to give a diastereomeric mixture of **21a** and **21b** along with recovered starting material **20a** (0.30 g, 30%).

Compound 21a: yield 761 mg (39%); mp 148–150 °C; R_f 0.45 (5% MeOH–CHCl₃); IR (KBr) 3410, 3160, 1690, 1675, 1650, 1410, 1355, 1270, 1200, 785, 735 cm⁻¹; ^1H NMR (DMSO- d_6) δ 1.56 (d, J = 7.3 Hz, 3 H, SeCHCH₃), 2.31 (s, 3 H, CH₃C=O), 3.78 (q, J = 7.3 Hz, 1 H, SeCHCH₃), 4.12 (1/2 ABq, J = 17.9 Hz, 1 H, CHH'N), 4.34 (1/2 ABq, J = 17.9 Hz, 1 H, CHH'N), 7.23 (s, 1 H, OH), 7.28–7.37 (m, 3 H, ArH), 7.43–7.55 (m, 2 H, ArH), 9.09 (s, 1 H, NH); ^{13}C NMR (DMSO- d_6) 15.59 (SeCHCH₃), 26.53 (CH₃C=O), 46.04 (CH₂N), 46.80 (SeCHCH₃), 84.32 (NC(OH)C(O)), 127.34, 129.00, 129.58, 132.99 (C₆H₅), 165.16, 168.30 (2 \times NC=O), 171.58 (CH₃C=O) ppm; the ^{13}C NMR assignments were confirmed by the corresponding APT experiment; MS (EI) m/e (rel intensity) 358 (4), 356 (21), 354 (9), 353 (4), 352 (4), 314 (72), 312 (88), 310 (55), 257 (85), 255 (55), 199 (59), 185 (100), 183 (85), 182 (61), 181 (64), 157 (97), 155 (83), 154 (67).

Anal. Calcd for C₁₄H₁₆N₂O₄Se: C, 47.33; H, 4.54; N, 7.89. Found: C, 47.43; H, 4.50; N, 7.83.

Compound 21b: yield 273 mg (14%); mp 147–150 °C; R_f 0.53 (5% MeOH–CHCl₃); IR (KBr) 3405, 3160, 1695, 1670, 1650, 1410, 1360, 1275, 780 cm⁻¹; ^1H NMR (CDCl₃) δ 1.49 (d, J = 7.0 Hz, 3 H, SeCHCH₃), 2.52 (s, 3 H, CH₃C=O), 3.95 (q, J = 7.0 Hz, 1 H, SeCHCH₃), 4.31 (1/2 ABq, J = 18.3 Hz, 1 H, CHH'N), 4.53 (1/2 ABq, J = 18.3 Hz, 1 H, CHH'N), 5.22 (br s, 1 H, OH), 7.24–7.29 (m, 3 H, ArH), 7.58–7.61 (m, 2 H, ArH), 8.00 (s, 1 H, NH); ^{13}C NMR (CDCl₃) 17.30 (SeCHCH₃), 26.82 (CH₃C=O), 46.35 (CH₂N), 46.84 (SeCHCH₃), 84.37 (NC(OH)C(O)), 127.17, 128.25, 129.24, 134.64 (C₆H₅), 166.66, 167.09 (2 \times NC=O), 171.59 (CH₃C=O) ppm; the ^{13}C NMR assignments were confirmed by the corresponding APT experiment; MS (EI) m/e (rel intensity) 358 (12), 356 (58), 354 (25), 353 (12), 352 (10), 199 (78), 185 (86), 183 (78), 182 (65), 181 (68), 157 (100), 155 (63).

Anal. Calcd for C₁₄H₁₆N₂O₄Se: C, 47.33; H, 4.54; N, 7.89. Found: C, 47.41; H, 4.56; N, 7.78.

Preparation of 1-Acetyl-3-methoxy-3-(1-(phenylselenenyl)ethyl)-2,5-piperazinedione (22a and 22b). A solution of **21a** (500 mg, 1.408 mmol) in MeOH (50 mL) and AcOH (10 mL) was stirred at rt (24 h). The solvent was removed in vacuo, and the residue was poured into H₂O (50 mL) and extracted with CHCl₃ (5 \times 50 mL). The organic layers were combined and washed with an aqueous saturated NaHCO₃ solution (3 \times 100 mL), an aqueous saturated NaCl solution (100 mL), and dried. The solvent was evaporated in vacuo, and the residue was purified by PTLC (5% MeOH–CHCl₃) to give a diastereomeric mixture of **22a** and **22b** along with recovered **21a** (0.23 g, 47%).

Compound 22a: yield 13 mg as an oil (3%); R_f 0.92 (5% MeOH–CHCl₃); IR (CHCl₃) 3310, 1715, 1700, 1685, 1425, 1365, 1250, 1210, 750 cm⁻¹; ^1H NMR (CDCl₃) δ 1.30 (d, J = 7.1 Hz, 3 H, CHCH₃), 2.61 (s, 3 H, CH₃C=O), 3.19 (s, 3 H, OCH₃), 4.12 (q, J = 7.1 Hz, 1 H, CHCH₃), 4.26 (1/2 ABq, J = 18.6 Hz, 1 H, CHH'N), 4.59 (1/2 ABq, J = 18.6 Hz, 1 H, CHH'N), 7.06 (s, 1 H, NH), 7.26–7.35 (m, 3 H, ArH), 7.62–7.65 (m, 2 H, ArH); ^{13}C NMR (CDCl₃) 17.02 (CHCH₃), 27.27 (CH₃C=O), 44.08 (CHCH₃), 46.43 (CH₂N), 51.73 (OCH₃), 89.68 (NC(OCH₃)C(O)), 127.40, 128.55, 129.14, 136.00 (C₆H₅), 165.37, 165.97 (2 \times NC=O), 171.70 (C-

H₃C=O) ppm; the ^{13}C NMR assignments were confirmed by the corresponding APT experiment; MS (EI) m/e (rel intensity) 372 (3), 370 (17), 368 (7), 367 (3), 366 (3), 185 (100), 183 (51), 157 (47).

Anal. Calcd for C₁₅H₁₆N₂O₄Se: C, 48.79; H, 4.91; N, 7.59. Found: C, 48.68; H, 4.76; N, 7.36.

Compound 22b: yield 205 mg (39%); mp 151–152 °C; R_f 0.78 (5% MeOH–CHCl₃); IR (KBr) 3180, 1705, 1675, 1425, 1360, 1245, 1205, 1075, 735 cm⁻¹; ^1H NMR (CDCl₃) δ 1.61 (d, J = 7.3 Hz, 3 H, CHCH₃), 2.37 (s, 3 H, CH₃C=O), 3.29 (s, 3 H, OCH₃), 3.91 (q, J = 7.3 Hz, 1 H, CHCH₃), 4.39 (1/2 ABq, J = 18.7 Hz, 1 H, CHH'N), 4.56 (1/2 ABq, J = 18.7 Hz, 1 H, CHH'N), 7.24–7.32 (m, 3 H, ArH), 7.46–7.49 (m, 2 H, ArH), 8.04 (s, 1 H, NH); ^{13}C NMR (CDCl₃) 15.22 (CHCH₃), 27.19 (CH₃C=O), 46.10 (CHCH₃), 46.54 (CH₂N), 52.16 (OCH₃), 89.70 (NC(OCH₃)C(O)), 128.13, 128.32, 129.20, 133.95 (C₆H₅), 165.88, 166.50 (2 \times NC=O), 171.98 (CH₃C=O) ppm; the ^{13}C NMR assignments were confirmed by the corresponding APT experiment; MS (EI) m/e (rel intensity) 372 (3), 370 (17), 368 (8), 367 (2), 366 (3), 185 (100), 183 (47).

Anal. Calcd for C₁₅H₁₆N₂O₄Se: C, 48.79; H, 4.91; N, 7.59. Found: C, 48.43; H, 4.71; N, 7.32.

Preparation of 1-Acetyl-3-methoxy-3-vinyl-2,5-piperazinedione (23). To a dry CH₂Cl₂ solution (20 mL) of **22a** and **22b** (120 mg, 0.325 mmol) was bubbled O₃ through the reaction at -20 °C until the solution became blue in color (1 min). The excess O₃ was then purged from the reaction system by bubbling O₂ through the solution (15 min). During this time the reaction was monitored for the presence of O₃ using an aqueous 2% KI indicator solution. The solvent was removed in vacuo, and the residue was purified by PTLC (5% MeOH–CHCl₃) to give **23** as an oil: yield 61 mg (89%); R_f 0.55 (5% MeOH–CHCl₃); IR (KBr) 3320, 2980, 1710, 1695, 1680, 1445, 1205, 1075 cm⁻¹; ^1H NMR (CDCl₃) δ 2.59 (s, 3 H, CH₃C=O), 3.30 (s, 3 H, OCH₃), 4.22 (1/2 ABq, J = 18.0 Hz, 1 H, CHH'N), 4.72 (1/2 ABq, J = 18.0 Hz, 1 H, CHH'N), 5.61 (d, J = 17.7 Hz, 1 H, CHH'=CH), 5.69 (d, J = 10.8 Hz, 1 H, CHH'=CH), 6.12 (dd, J = 10.8, 17.7 Hz, 1 H, CH₂=CH), 8.64 (s, 1 H, NH); ^{13}C NMR (CDCl₃) 26.97 (CH₃C=O), 45.90 (CH₂N), 50.64 (OCH₃), 85.86 (NC(OCH₃)C(O)), 120.70 (CH₂=CH), 132.15 (CH₂=CH), 165.35, 168.69 (2 \times NC=O), 171.67 (CH₃C=O) ppm; MS (EI) m/e (rel intensity) 169 (15), 139 (40), 127 (83), 112 (100).

Anal. Calcd for C₉H₁₂N₂O₄: C, 50.96; H, 5.66; N, 13.21. Found: C, 50.71; H, 5.67; N, 12.92.

Use of NMR Spectroscopy To Monitor the Preparation of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) from 1-Acetyl-3-methoxy-3-vinyl-2,5-piperazinedione (23). A solution of **23** (10 mg, 0.047 mmol) in THF- d_8 (0.10 mL) and DCl–D₂O (pD 3.96, 0.50 mL) was prepared and then monitored by ^1H and ^{13}C NMR spectroscopy. At the onset of the reaction the following NMR signals were observed: ^1H NMR (D₂O–THF- d_8) δ 2.43 (s, 3 H, CH₃C=O), 3.21 (s, 3 H, OCH₃), 4.24 (1/2 ABq, J = 18.0 Hz, 1 H, CHH'N), 4.36 (1/2 ABq, J = 18.0 Hz, 1 H, CHH'N), 5.48 (d, J = 17.7 Hz, 1 H, CHH'=CH), 5.52 (d, J = 10.8 Hz, 1 H, CHH'=CH), 5.88 (dd, J = 10.8, 17.7 Hz, 1 H, CH₂=CH); ^{13}C NMR (D₂O–THF- d_8) 26.79 (CH₃C=O), 46.34 (CH₂N), 51.46 (OCH₃), 87.19 (NC(OCH₃)C(O)), 121.36 (CH₂=CH), 133.22 (CH₂=CH), 167.02, 169.20 (2 \times NC=O), 174.18 (CH₃C=O) ppm. After 35 h, the signals corresponding to the starting material **23** were significantly reduced, and the following new set of signals attributed to **7** were observed: ^1H NMR (D₂O–THF- d_8) δ 2.43 (s, 3 H, CH₃C=O), 3.22 (s, 3 H, CH₃OD), 4.31 (s, CH₂N), 5.42 (d, J = 10.8 Hz, 1 H, CHH'=CH), 5.50 (d, J = 17.7 Hz, 1 H, CHH'=CH), 5.93 (dd, J = 10.8, 17.7 Hz, 1 H, CH₂=CH); ^{13}C NMR (D₂O–THF- d_8) 26.69 (CH₃C=O), 46.41 (CH₂N), 49.27 (CH₃OD), 82.22 (NC(OH)C(O)), 119.72 (CH₂=CH), 134.61 (CH₂=CH), 168.58, 168.74 (2 \times NC=O), 174.36 (CH₃C=O) ppm. The relative intensity of the two sets of peaks (**23** versus **7**) was approximately 1:1. After 96 h, the signals corresponding to the starting material **23** completely disappeared, and only the signals which corresponded to the title compound **7** and methanol- d_1 were detected: ^1H NMR (D₂O–THF- d_8) δ 2.44 (s, 3 H, CH₃C=O), 3.22 (s, 3 H, CH₃OD), 4.33 (s, CH₂N), 5.49 (d, J = 10.8 Hz, 1 H, CHH'=CH), 5.56 (d, J = 17.7 Hz, 1 H, CHH'=CH), 6.01 (dd, J = 10.8, 17.7 Hz, 1 H, CH₂=CH); ^{13}C NMR (CDCl₃) 26.70 (CH₃C=O), 46.39 (CH₂N), 49.26 (CH₃OD), 82.21 (NC(OH)C(O)), 119.76 (CH₂=CH), 134.58 (CH₂=CH), 168.54, 168.75 (2 \times NC=O), 174.48 (CH₃C=O) ppm.

Mass Spectral Identification and Isolation of Crude 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7). A solution of 23 (50 mg, 0.236 mmol) in a THF- d_8 -D₂O-CD₃CO₂D mixture ("pD" 2.0) was prepared and permitted to stand at rt (16 h). The ¹H and ¹³C NMR spectra indicated that the reaction was complete: ¹H NMR (THF- d_8 -D₂O-CD₃CO₂D) δ 2.44 (s, 3 H, CH₃C=O), 3.36 (s, 3 H, CH₃OD), 4.31 (s, CH₂N), 5.42 (d, J = 10.5 Hz, 1 H, CHH=CH), 5.50 (d, J = 17.2 Hz, 1 H, CHH=CH), 5.94 (dd, J = 10.5, 17.2 Hz, 1 H, CH₂=CH); ¹³C NMR (THF- d_8 -D₂O-CD₃CO₂D) 26.83 (CH₃C=O), 46.23 (CH₂N), 49.43 (CH₃OD), 82.50 (NC(OH)C(O)), 119.74 (CH₂=CH), 134.98 (CH₂=CH), 168.59, 169.14 (2 \times NC=O), 174.40 (CH₃C=O) ppm; MS (+FAB) 199 ([M + 1]⁺, M = C₈H₁₀N₂O₄), 200 ([M + 1]⁺, M = C₈H₉DN₂O₄), 201 ([M + 1]⁺, M = C₈H₈D₂N₂O₄); M_r (+FAB) 200.07786 (calcd for C₈H₉DN₂O₄, [M + 1]⁺ 200.07816). The solution was divided into two equal portions. The first portion was neutralized with an aqueous 0.01 N NaOH solution (final "pH" 6.9). The solvent was removed in vacuo (35 °C), and the residue was triturated with DMF- d_7 and filtered through glass wool. The filtrate exhibited the following NMR spectroscopic properties which have been attributed to 24: ¹H NMR (DMF- d_7) δ 1.83 (s, 3 H, CH₃C=O), 3.60 (d, J = 5.8 Hz, 2 H, CH₂NH), 7.01 (br s, 1 H, HH'NC=O), 7.32 (br s, 1 H, HH'NC=O), 8.02 (t, J = 5.8 Hz, 1 H, CH₂NH); ¹³C NMR (DMSO- d_6) 22.57 (CH₃C=O), 41.98 (CH₂N), 169.82, 171.43 (2 \times NC=O) ppm. Addition of authentic 24²⁵ to the NMR sample led to an increase in all the peaks attributed to 24, and no additional peaks were detected in either the ¹H or ¹³C NMR spectrum. To the second half of the reaction, was continuously added H₂O (10 mL), and the resulting azeotrope was evaporated in vacuo. This procedure provided a crude sample of 7 as an oil (~70% pure): ¹H NMR (DMSO- d_6) δ 2.38 (s, 3 H, CH₃C=O), 4.12 (1/2 ABq, J = 8.2 Hz, CHH'N), 4.31 (1/2 ABq, J = 8.2 Hz, CHH'N), 5.36 (d, J = 10.5 Hz, 1 H, CHH=CH), 5.48 (d, J = 17.2 Hz, 1 H, CHH=CH), 5.97 (dd, J = 10.5, 17.2 Hz, 1 H, CH₂=CH), 9.20 (s, 1 H, NH); ¹³C NMR (DMSO- d_6) 26.90 (CH₃C=O), 46.23 (CH₂N), 81.88 (NC(OH)C(O)), 117.39 (CH₂=CH), 135.78 (C-H₂=CH), 166.85, 167.77 (2 \times NC=O), 171.96 (CH₃C=O) ppm.

Use of ¹H NMR Spectroscopy to Monitor the Hydrolysis of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8). Solutions of 8 (10 mg, 0.055 mmol) in DCl-D₂O (1 mL, initial pD 4.36, 3.11, 2.31, and 1.08) were prepared and then monitored by ¹H NMR spectroscopy. No spectral changes were noted in the pD 4.36, 3.11, and 2.31 experiments after 24 h at rt, 12 h at 50 °C, and 1.5 h at 100 °C. Correspondingly, the pD 1.08 experiment exhibited no significant changes in the ¹H NMR spectrum at rt (24 h) and at 50 °C (12 h), but after the solution was maintained at 100 °C (1.5 h), signals corresponding to 8 were no longer evident and were replaced by the following resonances: ¹H NMR (D₂O) δ 2.64 (t, J = 5.7 Hz, 2 H), 3.80 (t, J = 5.7 Hz, 2 H), 3.87 (s, 2 H). A large peak was also observed at δ 4.80 and has been assigned to the residual H₂O present in the NMR sample. The signal at δ 3.87 has been attributed to 26. Addition of an authentic sample of 26²⁵ to the pD 1.08 solution led to an increase in the δ 3.87 peak.

Hydrolysis of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8). An aqueous 0.1 N HCl solution (1 mL) of 8 (30 mg, 0.165 mmol) was heated at 100 °C (1.5 h). The reaction mixture was cooled to rt and purified by PTLC (25% MeOH-CHCl₃) to obtain 25²⁷ (13 mg, 56%) and 26 (6 mg, 33%). Addition of an authentic sample of 26²⁵ to the NMR sample containing the product led to increases in the δ 3.86 (¹H NMR) and 42.89 and 171.81 (¹³C NMR) peaks. Correspondingly, addition of glycine led to new signals at δ 3.54 (¹H NMR) and 43.91 and 174.96 (¹³C NMR).

3-Hydroxy-4-(2-hydroxyethyl)-2(5H)-furanone (25):²⁷ R_f 0.70 (25% MeOH-CHCl₃); ¹H NMR (DMSO- d_6) δ 2.40 (t, J = 6.0 Hz, 2 H, HOCH₂CH₂), 3.57 (t, J = 6.0 Hz, 2 H, HOCH₂CH₂), 4.62 (s, 2 H, CH₂OC=O); ¹³C NMR (DMSO- d_6) 28.73 (HOCH₂CH₂), 59.15 (HOCH₂CH₂), 69.17 (CH₂OC=O), 129.84 (C=C-CH₂), 140.46 (C=CCH₂), 172.33 (C=O) ppm.

Glycinamide hydrochloride (26): R_f 0.15 (25% MeOH-CHCl₃); ¹H NMR (D₂O) δ 3.86 (s, 2 H, CH₂); ¹³C NMR (D₂O) 42.89 (CH₂), 171.81 (C=O) ppm.

Acetylation of 3-Hydroxy-4-(2-hydroxyethyl)-2(5H)-furanone (25). Preparation of 3-Acetoxy-4-(2-acetoxyethyl)-2(5H)-furanone (27).²⁷ A solution of 25 (10 mg, 0.069 mmol) in dry pyridine (1 mL) and Ac₂O (1 mL) was stirred at

rt under N₂ (16 h). The solvent was removed in vacuo, and the crude material was purified by PTLC (three developments) to give 27 (9 mg, 58%) as an oil: R_f 0.25 (CHCl₃); ¹H NMR (CDCl₃) δ 2.07 (s, 3 H, CH₃C=O), 2.31 (s, 3 H, CH₃C=O), 2.72 (t, J = 6.0 Hz, 2 H, OCH₂CH₂), 4.26 (t, J = 6.0 Hz, 2 H, OCH₂CH₂), 4.85 (s, 2 H, CH₂OC=O); ¹³C NMR (CDCl₃) 20.24 (CH₃C=O), 20.79 (CH₃C=O), 25.44 (OCH₂CH₂), 60.60 (OCH₂CH₂), 69.31 (CH₂OC=O), 135.35 (C=CCH₂), 146.08 (C=CCH₂), 166.98 (CH₃C=O), 167.10 (CH₃C=O), 170.56 (OC=O) ppm.

General Procedure for the Attempted Alcoholysis of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8). A solution of 8 (10 mg, 0.055 mmol) in anhydrous alcoholic 1 N H₂SO₄ (1 mL) was heated to reflux (1.5 h) under N₂. The solvent was removed in vacuo, and the residue was directly examined by ¹H NMR spectroscopy. Use of MeOH (65 °C) and PrOH (98 °C) in this procedure led to the recovery of only 8.

Attempted Propanolysis of Bicyclomycin (1). A solution of 1 (50 mg, 0.166 mmol) in anhydrous 1-propanolic 1 N H₂SO₄ (5 mL) was heated to reflux (1.5 h) under N₂. The solvent was removed in vacuo, and the residue was added to H₂O (5 mL) and extracted with EtOAc (5 \times 10 mL). The organic layers were combined, dried, and concentrated in vacuo. The residue was purified by PTLC (10% MeOH-CHCl₃) to give a 1:1 mixture²⁷ of rearranged products 34a and 34b: yield 12 mg (26%); mp 238–240 °C (lit.⁶ mp 238–242 °C); R_f 0.50 (10% MeOH-CHCl₃); ¹H NMR (DMSO- d_6) δ 1.22 (s, 3 H, CH₃), 2.68–2.75 (m, 2 H, C(3')H₂), 3.64 and 3.67 (2d, J = 9.0 Hz, 1 H, C(2)HH'), 3.81 and 3.84 (2d, J = 9.0 Hz, 1 H, C(2)HH'), 3.93 and 3.95 (2d, J = 7.1 Hz, 1 H, C(2')HH'), 4.00 and 4.04 (2d, J = 7.1 Hz, 1 H, C(2')HH'), 4.28 and 4.31 (2d, J = 6.0 Hz, 1 H, C(4)H), 5.07 (s, 1 H, C(3)OH), 5.27 and 5.32 (2s, 2 H, C=CH₂), 5.74 and 5.77 (2d, J = 6.0 Hz, 1 H, C(4)OH), 7.45 and 7.52 (2s, 1 H, NH), 9.10 and 9.13 (2s, 1 H, NH).

Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with Benzyl Mercaptan under Acidic Conditions. A solution of 8 (30 mg, 0.165 mmol) and benzyl mercaptan (100 mg, 0.805 mmol) in 25% THF-H₂O (5 mL, "pH" 1) was heated to reflux (4 h, 80 °C). During the reaction the concentration of benzyl mercaptan was determined at 30-min intervals with Ellman's reagent,³⁹ 5,5'-dithiobis(2-nitrobenzoic acid), to ensure that dimerization of benzyl mercaptan had not proceeded to an appreciable extent. After 4 h, greater than 1.7 equiv of benzyl mercaptan remained. The solvent was removed in vacuo, and the residue was purified by PTLC (10% MeOH-CHCl₃) to give 25 (11 mg, 46%) and glycinamide hydrochloride (26) (6 mg, 33%).

3-Hydroxy-4-(2-hydroxyethyl)-2(5H)-furanone (25):²⁷ R_f 0.70 (25% MeOH-CHCl₃); ¹H NMR (DMSO- d_6) δ 2.39 (t, J = 6.0 Hz, 2 H, HOCH₂CH₂), 3.57 (t, J = 6.0 Hz, 2 H, HOCH₂CH₂), 4.62 (s, 2 H, CH₂OC=O).

Glycinamide hydrochloride (26): R_f 0.15 (25% MeOH-CHCl₃); ¹H NMR (D₂O) δ 3.87 (s, 2 H, CH₂).

Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with EtSH at Near-Neutral "pH". To a tetrahydrofuran-aqueous Tris buffer solution (3:1, 1 mL, "pH" 8.0) were added 8 (5 mg, 0.027 mmol) and EtSH (27 mg, 0.440 mmol) (final "pH" 8.1). No change in the reaction was noted by TLC analysis after stirring at rt (96 h). The solvent was evaporated in vacuo, and the residue was purified by PTLC (10% MeOH-CHCl₃) to recover 8 (3.6 mg, 72%): ¹H NMR (DMSO- d_6) δ 2.58–2.71 (m, 2 H, OCH₂CH₂), 3.73–3.92 (m, 4 H, OCH₂CH₂, CH₂N), 5.03 (s, 1 H, C=CHH'), 5.22 (s, 1 H, C=CHH'), 8.11, 8.86 (2s, 2 H, 2 \times NH).

Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with Benzyl Mercaptan at Near-Neutral "pH". To a THF-H₂O solution (3:1, 1 mL) were added 8 (5 mg, 0.027 mmol) and benzyl mercaptan (6.8 mg, 0.055 mmol). The "pH" was adjusted from 7.7 to 8.1 with dilute aqueous NaOH,

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and then the solution was stirred at rt (48 h). No change in the reaction was noted by TLC analysis. The solvent was removed in vacuo, and the residue was purified by PTLC (10% MeOH-CHCl₃) to recover **8** (2.9 mg, 58%): ¹H NMR (DMSO-*d*₆) δ 2.60–2.73 (m, 2 H, OCH₂CH₂), 3.76–3.94 (m, 4 H, OCH₂CH₂, CH₂N), 5.05 (s, 1 H, C=CHH'), 5.24 (s, 1 H, C=CHH'), 8.15, 8.88 (2s, 2 H, 2 × NH).

General Procedure for the Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with Imidazole. Solutions of **8** (5 mg, 0.027 mmol) and imidazole (3 mg, 0.044 mmol) in THF-H₂O (3:1, 1 mL, final "pH" 9.0, 10.2, and 11.0) were prepared and then monitored by TLC analysis. No changes in these reactions were noted after stirring at rt (48 h). In each case the solvent was removed in vacuo, and the residue was purified by PTLC (10% MeOH-CHCl₃) to recover **8** (2.5–3.3 mg, 50–66%) and imidazole (1.5–1.9 mg, 50–63%).

Compound 8: ¹H NMR (DMSO-*d*₆) δ 2.61–2.74 (m, 2 H, OCH₂CH₂), 3.76–3.95 (m, 4 H, OCH₂CH₂, CH₂N), 5.05 (s, 1 H, C=CHH'), 5.25 (s, 1 H, C=CHH'), 8.17, 8.89 (2s, 2 H, 2 × NH).

Imidazole: mp 88–90 °C (lit.⁴⁰ mp 89–91 °C); *R*_f 0.40 (10% MeOH-CHCl₃); ¹H NMR (CDCl₃) δ 7.13 (s, 2 H, NCHCHN), 7.72 (s, 1 H, NCHN), 11.57 (br s, 1 H, NH).

Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with NaSMe under Basic Conditions. To a solution of **8** (40 mg, 0.220 mmol) in 25% H₂O-THF (3 mL) was added NaSMe (36 mg, 0.513 mmol) in H₂O (1 mL) (final "pH" 12.5). The solution was stirred at rt (2 h) and then neutralized ("pH" 7.3) with 0.1 N aqueous HCl. The solvent was removed in vacuo, and the residue was purified by PTLC (15% MeOH-CHCl₃) to give **36** as an oil: yield 35 mg (69%); *R*_f 0.62 (15% MeOH-CHCl₃); IR (KBr) 3240, 3100, 2920, 1665, 1625, 1435, 1410, 1315, 1095, 1045, 905 cm⁻¹; ¹H NMR (acetone-*d*₆) δ 1.98 (s, 3 H, SCH₃), 2.55 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂), 3.91 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂), 3.93 (s, 2 H, CH₂S), 4.01 (s, 2 H, CH₂N); ¹³C NMR (acetone-*d*₆) 13.67 (SCH₃), 33.40 (CH₂CH₂C=C or SCH₂), 33.82 (CH₂CH₂C=C or SCH₂), 45.32 (CH₂N), 59.76 (OCH₂CH₂), 124.26 (CH₂C=C), 132.02 (CH₂C=C), 163.43 (C=O), 165.26 (C=O) ppm; MS (EI) *m/e* (rel intensity) 230 (98), 212 (32), 197 (44), 183 (100), 165 (65), 151 (34); *M*_r (EI) 230.07225 (calcd for C₉H₁₄N₂O₃S, 230.07249), 212.06230 (calcd for C₈H₁₂N₂O₂S, 212.06195), 197.03842 (calcd for C₈H₉N₂O₂S, 197.03847), 183.07635 (calcd for C₈H₁₁N₂O₃, 183.07697), 165.06509 (calcd for C₈H₉N₂O₂, 165.06640).

Acetylation of 3-(3-Hydroxy-1-((methylthio)methyl)propylidene)-2,5-piperazinedione (36). Preparation of 3-(3-Acetoxy-1-((methylthio)methyl)propylidene)-2,5-piperazinedione (37). A solution of **36** (23 mg, 0.100 mmol) in dry pyridine (1 mL) and Ac₂O (1 mL) was stirred at rt under N₂ (24 h). The solvent was removed in vacuo, and the residue was purified by PTLC (5% MeOH-CHCl₃) to give **37** as an oil: yield 16 mg (60%); *R*_f 0.64 (5% MeOH-CHCl₃); IR (CDCl₃) 2987, 2256, 1736, 1377, 1251, 1047, 906, 738, 653 cm⁻¹; ¹H NMR (CDCl₃) δ 2.04 (s, 3 H, SCH₃), 2.13 (CH₃C=O), 2.64 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂), 3.97 (s, 2 H, CH₂S), 4.09 (s, 2 H, CH₂N), 4.26 (t, *J* = 6.0 Hz, 2 H, OCH₂CH₂); ¹³C NMR (CDCl₃) 14.47 (SCH₃), 20.87 (CH₃C=O), 31.31 (OCH₂CH₂), 34.18 (CH₂S), 45.20 (CH₂N), 63.08 (OCH₂CH₂), 124.79 (CH₂C=C), 127.63 (CH₂C=C), 161.13 (C=CC=O), 164.14 (CH₂C=O), 171.64 (CH₃C=O) ppm; MS (EI) *m/e* (rel intensity) 272 (8), 230 (9), 212 (69), 183 (13), 165 (100), 137 (56), 108 (26); *M*_r (EI) 272.08295 (calcd for C₁₁H₁₆N₂O₄S, 272.08308), 212.06130 (calcd for C₉H₁₂N₂O₂S, 212.06195), 165.06618 (calcd for C₈H₉N₂O₂, 165.06640), 137.07173 (calcd for C₇H₉N₂O, 137.07149).

Treatment of 4-Methylene-1-oxa-6,9-diazaspiro[4.5]decane-7,10-dione (8) with NaSMe in Basic MeOH. A solution of **8** (40 mg, 0.220 mmol) and NaSMe (36 mg, 0.513 mmol) in 0.03 N NaOMe-MeOH (final "pH" 11.9) was stirred at rt under N₂ (24 h). The solution was neutralized ("pH" 7.1) with 0.1 N methanolic H₂SO₄. The solvent was removed in vacuo, and the residue was purified by PTLC (10% MeOH-CHCl₃) to recover **8** (33 mg, 83%): ¹H NMR (DMSO-*d*₆) δ 2.60–2.74 (m, 2 H, OCH₂CH₂), 3.76–3.95 (m, 4 H, OCH₂CH₂, CH₂N), 5.05 (s, 1 H, C=CHH'), 5.25 (s, 1 H, C=CHH'), 8.18, 8.90 (2s, 2 H, 2 × NH).

Treatment of Bicyclomycin (1) with NaSMe in Basic MeOH. A solution of **1** (50 mg, 0.166 mmol) and NaSMe (36 mg, 0.513 mmol) in 0.03 N NaOMe-MeOH (final "pH" 11.9) was

stirred at rt under N₂ (2 h). The solution was neutralized ("pH" 7.0) with 0.1 N methanolic H₂SO₄. The solvent was removed in vacuo, and the residue was purified by PTLC (20% MeOH-CHCl₃) to give a diastereomeric mixture^{7a,15} of **38a** (27 mg, 46%) and **38b** (12 mg, 20%) as a semisolid and an unidentified minor product (5 mg).

Compound 38a: *R*_f 0.45 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 1.17 (s, 3 H, C(2')CH₃), 1.65–1.89 (m, 1 H, C(4)HH'), 2.01 (s, 3 H, SCH₃), 2.08–2.20 (m, 1 H, C(4)HH'), 2.21–2.40 (m, 2 H, C(5)H, C(5a)HH'), 2.57–2.73 (m, 1 H, C(5a)HH'), 3.57–3.81 (m, 4 H, C(1')H, C(3')H₂, C(3)HH'), 3.86–4.00 (m, 1 H, C(3)HH'), 4.79 (s, 1 H, C(1')OH), 6.25 (s, 1 H, C(2')OH), 6.49 (s, 1 H, C(6)OH), 7.11, 7.20 (2s, 2 H, N(10)H₂), 8.04 (s, 1 H, N(8)H); ¹³C NMR (DMSO-*d*₆) 15.05 (SCH₃), 23.65 (C(2')CH₃), 29.73 (C(5a)), 32.96 (C(4)), 46.43 (C(5)), 66.81 (C(3)), 74.83 (C(2')), 76.02 (C(3')), 78.96 (C(1')), 88.15 (C(1)), 101.86 (C(6)), 169.49 (C(7) or C(9)), 171.89 (C(7) or C(9)) ppm; MS (+FAB) *m/e* (rel intensity) 373 [M + Na⁺] (100), 351 [M + H⁺] (71), 333 (77), 316 (40); *M*_r (+FAB) 373.10405 (calcd for C₁₃H₂₂NaN₂O₇S, 373.10454).

Compound 38b: *R*_f 0.41 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 1.17 (s, 3 H, C(2')CH₃), 1.65–1.89 (m, 1 H, C(4)HH'), 2.01 (s, 3 H, SCH₃), 2.07–2.20 (m, 1 H, C(4)HH'), 2.24–2.43 (m, 2 H, C(5)H, C(5a)HH'), 2.57–2.75 (m, 1 H, C(5a)HH'), 3.40–3.82 (m, 4 H, C(1')H, C(3')H₂, C(3)HH'), 3.89–4.00 (m, 1 H, C(3)HH'), 4.59 (s, 1 H, C(1')OH), 5.45 (s, 1 H, C(2')OH), 6.59 (s, 1 H, C(6)OH), 7.60, 7.83 (2s, 2 H, N(10)H₂), 8.71 (s, 1 H, N(8)H); ¹³C NMR (DMSO-*d*₆) 15.00 (SCH₃), 23.58 (C(2')CH₃), 29.49 (C(5a)), 32.88 (C(4)), 46.41 (C(5)), 66.85 (C(3)), 74.90 (C(2')), 76.04 (C(3')), 78.93 (C(1')), 88.09 (C(1)), 101.89 (C(6)), 169.47 (C(7) or C(9)), 171.85 (C(7) or C(9)) ppm; MS (+FAB) *m/e* (rel intensity) 373 [M + Na⁺] (100), 351 [M + H⁺] (68), 333 (100); *M*_r (+FAB) 373.10405 (calcd for C₁₃H₂₂NaN₂O₇S, 373.10454).

Treatment of Compounds 34a and 34b with NaSMe under Basic Conditions. A THF-H₂O mixture ("pH" 12.4) containing **34a** and **34b** (40 mg, 0.14 mmol) and NaSMe (20 mg, 0.28 mmol) was stirred at rt (24 h) under Ar. The solution was neutralized with aqueous 0.1 N HCl, and the solvent was removed in vacuo. The residue was subjected to preparative TLC (15% MeOH-CHCl₃) to give **34a** (**34b**)⁶ as the major component: *R*_f 0.55 (15% MeOH-CHCl₃); ¹H NMR (CD₃OD) δ 1.33 (s, 3 H, C(3)CH₃), 2.76–2.95 (m, 2 H, C(3')H₂), 3.82 (d, *J* = 9.3 Hz, 1 H, C(2)HH'), 3.95 (d, *J* = 9.3 Hz, 1 H, C(2)HH'), 4.05–4.15 (m, 2 H, C(2')H₂), 4.41 (s, 1 H, C(4)H), 5.24–5.26 (m, 1 H, C(4')=CHH'), 5.33–5.35 (m, 1 H, C(4')=CHH'); ¹³C NMR (CD₃OD) 21.18 (C(3)CH₃), 33.21 (C(3')), 68.31 (C(2')), 77.56 (C(4)), 78.75 (C(3)), 78.83 (C(2)), 90.15 (C(5)), 111.62 (C(4')=CH₂), 150.43 (C(4')), 168.56 (C=O), 169.10 (C=O). The remaining quaternary carbon was not detected. Inspection of the ¹³C spectrum indicated that the product existed as a single diastereoisomer.

Hydrolysis of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) under Acidic Conditions. Compound **7** (15 mg, 0.076 mmol) was prepared at "pD" 2.0 in situ (THF-*d*₈-D₂O-CD₃CO₂D), and then the "pD" of the solution was adjusted to 1.0 with 1.0 N DCl-D₂O. The resulting solution was permitted to stand at rt (16 h). TLC analysis prior to workup indicated that the reaction was complete and no other significant product was noted other than **41** that migrated beyond the origin. The solution was neutralized ("pD" 7.0), and then the solvent was removed in vacuo. The residue was purified by PTLC (20% MeOH-CHCl₃) to give **41**: yield 6 mg (51%); mp 239–241 °C; *R*_f 0.25 (20% MeOH-CHCl₃); IR (KBr) 3305, 3130, 2930, 1670, 1625, 1435, 1095 cm⁻¹; ¹H NMR (DMSO-*d*₆) δ 3.91 (s, 2 H, CH₂N), 4.17 (d, *J* = 5.7 Hz, 2 H, CHCH₂O), 5.42 (s, 1 H, OH), 5.71 (t, *J* = 5.7 Hz, 1 H, CHCH₂O), 8.15, 8.17 (2s, 2 H, 2 × NH); ¹³C NMR (DMSO-*d*₆) 46.47 (CH₂N), 56.81 (CHCH₂O), 115.68 (C=CHCH₂), 126.83 (C=CHCH₂), 158.48, 163.48 (2 × NC=O) ppm; MS (+CI) *m/e* (rel intensity) 157 (100), 139 (72), 107 (88); *M*_r (EI) 156.05849 (calcd for C₈H₈N₂O₃, 156.05350).

Hydrolysis of 1-Acetyl-3-methoxy-3-vinyl-2,5-piperazinedione (23) under Acidic Conditions. Compound **23** (20 mg, 0.094 mmol) was dissolved in 25% H₂O-THF (1 mL), and then the "pH" was adjusted to 1.0 with an aqueous 1.0 N HCl solution. The resulting solution was permitted to stand at rt (16 h). TLC analysis prior to workup indicated that the reaction was complete, and no other significant product was noted other than **41** that migrated beyond the origin. The solution was neutralized

("pH" 7.0), and the solvent was removed in vacuo. The residue was purified by PTLC (20% MeOH-CHCl₃) to give 41: yield 5 mg (34%); mp 239–241 °C; *R_f* 0.25 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 3.93 (s, 2 H, CH₂N), 4.20 (d, *J* = 5.7 Hz, 2 H, CHCH₂O), 5.51 (s, 1 H, OH), 5.72 (t, *J* = 5.7 Hz, 1 H, CHCH₂O), 8.13, 8.17 (2 s, 2 H, 2 × NH); ¹³C NMR (DMSO-*d*₆) 46.09 (CH₂N), 56.89 (CHCH₂O), 115.74 (C=CHCH₂), 126.86 (C=CHCH₂), 159.00, 163.46 (2 × NC=O) ppm.

Methanolysis of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) under Acidic Conditions. A solution of crude 7 (30 mg, 0.153 mmol) in methanolic H₂SO₄ ("pH" 1.0) was permitted to stand at rt (12 h). The solution was neutralized to "pH" 6.8. The solvent was removed in vacuo, and the residue was purified by PTLC (30% acetone-chloroform) to give 23 (17 mg, 53%) and a small amount of product (~0.6 mg, 2%) which has been tentatively assigned as a 2:1 mixture of (*Z*)-42a and (*E*)-42b, 1-acetyl-3-(2-methoxyethylidene)-2,5-piperazinedione.

1-Acetyl-3-methoxy-3-vinyl-2,5-piperazinedione (23): ¹H NMR (CDCl₃) δ 2.59 (s, 3 H, CH₃C=O), 3.31 (s, 3 H, OCH₃), 4.19 (1/2 ABq, *J* = 18.0 Hz, 1 H, CHH'N), 4.71 (1/2 ABq, *J* = 18.0 Hz, 1 H, CHH'N), 5.61 (d, *J* = 17.7 Hz, 1 H, CHH'=CH), 5.64 (d, *J* = 10.8 Hz, 1 H, CHH'=CH), 6.11 (dd, *J* = 10.8, 17.7 Hz, 1 H, CH₂=CH), 8.69 (s, 1 H, NH).

(*Z*)-1-Acetyl-3-(2-methoxyethylidene)-2,5-piperazinedione (42a): ¹H NMR (DMSO-*d*₆) δ 2.50 (s, 3 H, CH₃C=O), 3.94 (s, 2 H, CH₂N), 4.09 (d, *J* = 6.5 Hz, 2 H, CHCH₂O), 5.71 (t, *J* = 6.5 Hz, 1 H, CHCH₂O), 9.95 (s, 1 H, NH).

(*E*)-1-Acetyl-3-(2-methoxyethylidene)-2,5-piperazinedione (42b): ¹H NMR (DMSO-*d*₆) δ 2.50 (s, 3 H, CH₃C=O), 3.87 (s, 2 H, CH₂N), 4.38 (d, *J* = 6.5 Hz, 2 H, CHCH₂O), 5.44 (t, *J* = 6.5 Hz, 1 H, CHCH₂O), 10.25 (s, 1 H, NH).

Hydrolysis of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) at "pD" 6.0. Compound 7 (10 mg, 0.051 mmol) was prepared in situ at "pD" 2.0 in THF-*d*₈-D₂O-CD₃CO₂D, and the "pD" was adjusted to 6.0 with 1.0 N NaOD-D₂O. The solution was permitted to stand at rt (3 h). TLC analysis prior to workup indicated that the reaction was complete, and no other significant product was noted other than 41 that migrated beyond the origin. The solution was neutralized to "pD" 7.0, and then the solvent was removed in vacuo. The residue was purified by PTLC (20% MeOH-CHCl₃) to give 41: yield 3 mg (38%); mp 238–241 °C; *R_f* 0.25 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 3.90 (s, 2 H, CH₂N), 4.19 (d, *J* = 5.7 Hz, 2 H, CHCH₂O), 5.43 (s, 1 H, OH), 5.73 (t, *J* = 5.7 Hz, 1 H, CHCH₂O), 8.07, 8.14 (2 s, 2 H, 2 × NH).

Hydrolysis of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) at "pD" 8.8. Compound 7 (15 mg, 0.076 mmol) was prepared in situ at "pD" 2.0 in THF-*d*₈-D₂O-CD₃CO₂D and the "pD" was adjusted to 8.8 with 1.0 N NaOD-D₂O. The resulting solution was permitted to stand at rt (30 min). The solution was neutralized to "pD" 7.0 with DCl-D₂O. The solvent was removed in vacuo, and the residue was purified by PTLC (20% MeOH-CHCl₃) to give 41 (2 mg, 13%) and 24 (4 mg, 45%).

3-(2-Hydroxyethylidene)-2,5-piperazinedione (41): mp

238–241 °C; *R_f* 0.25 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 3.90 (s, 2 H, CH₂N), 4.19 (d, *J* = 5.7 Hz, 2 H, CHCH₂O), 5.43 (s, 1 H, OH), 5.73 (t, *J* = 5.7 Hz, 1 H, CHCH₂O), 8.07, 8.14 (2 s, 2 H, 2 × NH).

***N*_α-Acetylglycinamide (24):** *R_f* 0.05 (10% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 1.87 (s, 3 H, CH₃C=O), 3.57 (d, *J* = 5.8 Hz, 2 H, CH₂NH), 7.00 (br s, 1 H, HH'NC=O), 7.36 (br s, 1 H, HH'NC=O), 8.06 (t, *J* = 5.8 Hz, 1 H, CH₂NH). An authentic sample of 24²⁵ was added to the NMR sample, and no new additional peaks were observed.

Studies on the Stability of Bicyclomycin in Near-Neutral and Moderately Basic Solutions. Bicyclomycin (1) (10 mg, 0.03 mmol) was added to a buffered KD₂PO₄ (0.10 M) D₂O-THF-*d*₈ (2:1) solution (0.50 mL, "pD" 5.7), and then the solution was maintained at rt. The reaction was monitored by ¹H NMR spectroscopy. No detectable change was noted after 24 h.

Bicyclomycin (1) (10 mg, 0.03 mmol) was added to a buffered NaDCO₃ (0.10 M) D₂O-THF-*d*₈ (2:1) solution (0.50 mL, "pD" 10.2), and then the solution was maintained at rt. The reaction was monitored by ¹H NMR spectroscopy. After 24 h, approximately 10% of 1 had been converted into a new adduct as evidenced by the appearance of a new set of vinylic protons upfield from the corresponding signals in 1.

Treatment of 1-Acetyl-3-hydroxy-3-vinyl-2,5-piperazinedione (7) with NaSMe under Near-Neutral Conditions ("pD" 7.0). Compound 7 (10 mg, 0.051 mmol) was prepared at "pD" 2.0 in THF-*d*₈-D₂O-CD₃CO₂D in situ, the "pD" was adjusted to 4.9 with 1.0 N NaOD-D₂O, and then NaSMe (14 mg, 0.200 mmol) in a 3:1 mixture of THF-*d*₈-D₂O (0.5 mL) was added. The final "pD" of the resulting solution was adjusted to 7.0 with 0.1 N NaOD-D₂O. The solution was permitted to stand at rt (24 h). TLC analysis prior to workup indicated that the reaction was complete, and no other significant product was noted other than 41 that migrated beyond the origin. The solvent was removed in vacuo, and the residue was purified by PTLC (20% MeOH-CHCl₃) to give 41: yield 2 mg (19%); *R_f* 0.25 (20% MeOH-CHCl₃); ¹H NMR (DMSO-*d*₆) δ 3.92 (s, 2 H, CH₂N), 4.18 (d, *J* = 5.7 Hz, 2 H, CHCH₂O), 5.44 (s, 1 H, OH), 5.74 (t, *J* = 5.7 Hz, 1 H, CHCH₂O), 8.17, 8.25 (2 s, 2 H, 2 × NH).

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